

Ancient human genomes suggest three ancestral populations for present-day Europeans

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We sequenced the genomes of a ~7,000-year-old farmer from Germany and eight ~8,000-year-old hunter-gatherers from Luxembourg and Sweden. We analysed these and other ancient genomes^{1–4} with 2,345 contemporary humans to show that most present-day Europeans derive from at least three highly differentiated populations: west European hunter-gatherers, who contributed ancestry to all Europeans but not to Near Easterners; ancient north Eurasians related to Upper Palaeolithic Siberians³, who contributed to both Europeans and Near Easterners; and early European farmers, who were mainly of Near Eastern origin but also harboured west European hunter-gatherer related ancestry. We model these populations' deep relationships and show that early European farmers had ~44% ancestry from a 'basal Eurasian' population that split before the diversification of other non-African lineages.

Near Eastern migrants from Anatolia and the Levant are known to have played a major role in the introduction of agriculture to Europe, as ancient DNA indicates that early European farmers were distinct from European hunter-gatherers^{4,5} and close to present-day Near Easterners^{4,6}. However, modelling present-day Europeans as a mixture of these two ancestral populations⁴ does not account for the fact that Europeans are also admixed with a population related to Native Americans^{7,8}. To clarify the prehistory of Europe, we sequenced nine ancient genomes (Fig. 1 and Extended Data Fig. 1): 'Stuttgart' (19-fold coverage), a ~7,000-year-old skeleton found in Germany in the context of artefacts from the first widespread farming culture of central Europe, the Linearbandkeramik; 'Loschbour' (22-fold), an ~8,000-year-old skeleton from the Loschbour rock shelter in Luxembourg, discovered in the context of hunter-gatherer artefacts (Supplementary Information sections 1 and 2); and seven ~8,000-year-old samples (0.01–2.4-fold) from a hunter-gatherer burial in Motala, Sweden (the highest coverage individual was 'Motala12').

Sequence reads from all samples revealed >20% C→T and G→A deamination-derived mismatches at the ends of the molecules that are characteristic of ancient DNA^{9,10} (Supplementary Information section 3). We estimate nuclear contamination rates to be 0.3% for Stuttgart and 0.4% for Loschbour (Supplementary Information section 3), and mitochondrial (mtDNA) contamination rates to be 0.3% for Stuttgart, 0.4% for Loschbour, and 0.01–5% for the Motala individuals (Supplementary Information section 3). Stuttgart has mtDNA haplogroup T2, typical of Neolithic Europeans¹¹, and Loschbour and all Motala individuals have the U5 or U2 haplogroups, typical of hunter-gatherers^{5,9} (Supplementary Information section 4). Stuttgart is female, whereas Loschbour and five Motala individuals are male (Supplementary Information section 5) and belong to Y-chromosome haplogroup I, suggesting that this was common in pre-agricultural Europeans (Supplementary Information section 5).

We carried out large-scale sequencing of libraries prepared with uracil DNA glycosylase (UDG), which removes deaminated cytosines, thus reducing errors arising from ancient DNA damage (Supplementary Information section 3). The ancient individuals had indistinguishable levels of Neanderthal ancestry when compared to each other (~2%) and to present-day Eurasians (Supplementary Information section 6). The heterozygosity of Stuttgart (0.00074) is at the high end of present-day Europeans, whereas that of Loschbour (0.00048) is lower than in any present human populations (Supplementary Information section 2); this must

reflect a strong bottleneck in Loschbour's ancestors, as the genetic data show that he was not recently inbred (Extended Data Fig. 2). High copy numbers for the salivary amylase gene (*AMY1*) have been associated with a high starch diet¹²; our ancient genomes are consistent with the direction of this observation in that the Stuttgart farmer had the highest number of copies (16), whereas the ancient hunter-gatherers La Braña (from Iberia)², Motala12, and Loschbour had lower numbers (5, 6 and 13, respectively) (Supplementary Information section 7). We caution, however, that copy count in Loschbour is at the high end of present-day humans, showing that high copy counts of *AMY1* cannot be accounted for entirely by selection since the switch to agriculture. Both Loschbour and Stuttgart had dark hair (>99% probability); and Loschbour, like La Braña and Motala12, probably had blue or light coloured eyes (>75%) whereas Stuttgart probably had brown eyes (>99% probability) (Supplementary Information section 8). Neither Loschbour nor La Braña carries the skin-lightening allele in *SLC24A5* that is homozygous in Stuttgart and nearly fixed in Europeans today², but Motala12 carries at least one copy of the derived allele, showing that this allele was present in Europe before the advent of agriculture.

We compared the ancient genomes to 2,345 present-day humans from 203 populations genotyped at 594,924 autosomal single nucleotide polymorphisms (SNPs) with the Human Origins array⁸ (Supplementary Information section 9) (Extended Data Table 1). We used ADMIXTURE¹³ to identify 59 'west Eurasian' populations that cluster with Europe and the Near East (Supplementary Information section 9 and Extended Data Fig. 3). Principal component analysis (PCA)¹⁴ (Supplementary Information section 10) (Fig. 2) indicates a discontinuity between the Near East and Europe, with each showing north–south clines bridged only by a few populations of mainly Mediterranean origin. We projected¹⁵ the newly sequenced and previously published^{1–4} ancient genomes onto the first two principal components (PCs) (Fig. 2). Upper Palaeolithic hunter-gatherers³ from Siberia like the MA1 (Mal'ta) individual project at the northern end of the PCA, suggesting an 'ancient north Eurasian' (ANE) meta-population. European hunter-gatherers from Spain², Luxembourg, and Sweden⁴ fall beyond present-day Europeans in the direction of European differentiation from the Near East, and form a 'west European hunter-gatherer' (WHG) cluster including Loschbour and La Braña², and a 'Scandinavian hunter-gatherer' (SHG) cluster including the Motala individuals and ~5,000-year-old hunter-gatherers from the Pitted Ware Culture⁴. An 'early European farmer' (EEF) cluster includes Stuttgart, the ~5,300-year-old Tyrolean Iceman¹ and a ~5,000-year-old Swedish farmer⁴.

Patterns observed in PCA may be affected by sample composition (Supplementary Information section 10) and their interpretation in terms of admixture events is not straightforward, so we rely on formal analysis of *f* statistics⁸ to document mixture of at least three source populations in the ancestry of present Europeans. We began by computing all possible statistics of the form $f_3(\text{Test}, \text{Ref}_1, \text{Ref}_2)$ (Supplementary Information section 11), which if significantly negative show unambiguously⁸ that *Test* is admixed between populations anciently related to *Ref*₁ and *Ref*₂ (we choose *Ref*₁ and *Ref*₂ from 5 ancient and 192 present populations). The lowest *f*₃ statistics for Europeans are negative (93% are > 4 standard errors below 0), with most showing strong support for at least one

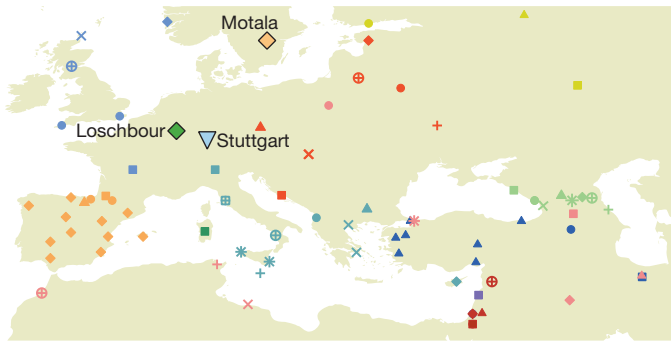


Figure 1 | Map of west Eurasian populations. Geographical locations of analysed samples, with colour coding matching the PCA (Fig. 2). We show all sampling locations for each population, which results in multiple points for some (for example, Spain).

ancient individual being one of the references (Supplementary Information section 11). Europeans almost always have their lowest f_3 with either (EEF, ANE) or (WHG, Near East) (Supplementary Information section 11, Table 1 and Extended Data Table 1), which would not be expected if there were just two ancient sources of ancestry (in which case the best references for all Europeans would be similar). The lowest f_3 statistic for Near Easterners always takes Stuttgart as one of the reference populations, consistent with a Near Eastern origin for Stuttgart's ancestors (Table 1). We also computed the statistic $f_4(Test, Stuttgart; MA1, Chimp)$, which measures whether MA1 shares more alleles with a *Test* population or with Stuttgart. This statistic is significantly positive (Extended Data Fig. 4 and Extended Data Table 1) if *Test* is nearly any present-day West Eurasian population, showing that MA1-related ancestry has increased since the time of early farmers like Stuttgart (the same statistic using Native Americans instead of MA1 has the same

sign but is smaller in magnitude (Extended Data Fig. 5), indicating that MA1 is a better surrogate than the Native Americans who were first used to document ANE ancestry in Europe^{7,8}). The analogous statistic $f_4(Test, Stuttgart; Loschbour, Chimp)$ is nearly always positive in Europeans and negative in Near Easterners, indicating that Europeans have more ancestry from populations related to Loschbour than do Near Easterners (Extended Data Fig. 4 and Extended Data Table 1). Extended Data Table 2 documents the robustness of key f_4 statistics by recomputing them using transversion polymorphisms not affected by ancient DNA damage, and also using whole-genome sequencing data not affected by SNP ascertainment bias. Extended Data Fig. 6 shows the geographic gradients in the degree of allele sharing of present-day West Eurasians (as measured by f_4 statistics) with Stuttgart (EEF), Loschbour (WHG) and MA1 (ANE).

To determine the minimum number of source populations needed to explain the data for many European populations taken together, we studied the matrix of all possible statistics of the form $f_4(Test_{base}, Test_i; O_{base}, O_j)$ (Supplementary Information section 12). $Test_{base}$ is a reference European population, $Test_i$ is the set of all other European *Test* populations, O_{base} is a reference outgroup, and O_j is the set of other outgroups (ancient DNA samples, Onge, Karitiana, and Mbuti). The rank of the (i, j) matrix reflects the minimum number of sources that contributed to the *Test* populations^{16,17}. For a pool of individuals from 23 *Test* populations representing most present-day European groups, this analysis rejects descent from just two sources ($P < 10^{-12}$ by a Hotelling *t*-test¹⁷). However, three source populations are consistent with the data after excluding the Spanish who have evidence for African admixture^{18–20} ($P = 0.019$, not significant after multiple-hypothesis correction), consistent with the results from ADMIXTURE (Supplementary Information section 9), PCA (Fig. 2 and Supplementary Information section 10) and *f* statistics (Extended Data Table 1, Extended Data Fig. 6, Supplementary Information sections 11 and 12). We caution that the finding of three sources could be consistent with a larger number of mixture events. Moreover, the source

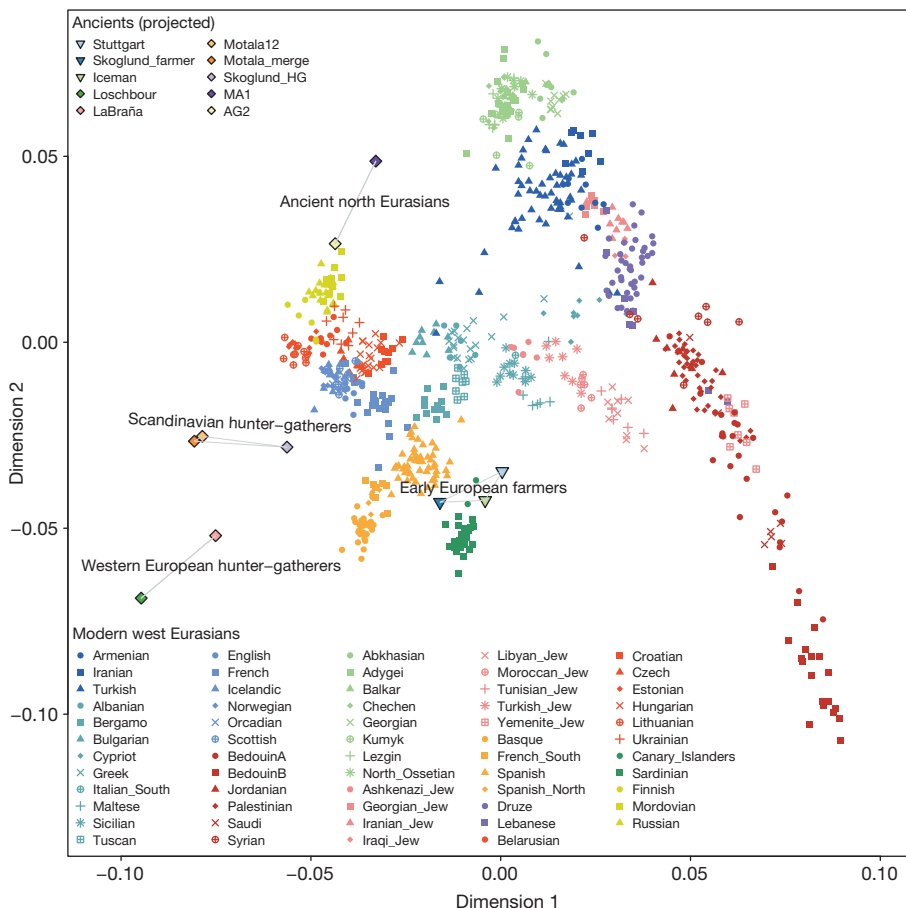


Figure 2 | Principal Component Analysis. PCA on all present-day west Eurasians, with ancient samples projected. European hunter-gatherers fall beyond present-day Europeans in the direction of European differentiation from the Near East. Stuttgart clusters with other Neolithic Europeans and present-day Sardinians. MA1 falls outside the variation of present-day west Eurasians in the direction of southern–northern differentiation along dimension 2.

Table 1 | Lowest f_3 statistics for each west Eurasian population

Ref_1	Ref_2	Target for which these two references give the lowest $f_3(X; Ref_1, Ref_2)$
WHG	EEF	Sardinian***
WHG	Near East	Basque, Belarusian, Czech, English, Estonian, Finnish, French_South, Icelandic, Lithuanian, Mordovian, Norwegian, Orcadian, Scottish, Spanish, Spanish_North, Ukrainian
WHG	Siberian	Russian
EEF	ANE	Abkhasian***, Albanian, Ashkenazi_Jew****, Bergamo, Bulgarian, Chechen****, Croatian, Cypriot****, Druze**, French, Greek, Hungarian, Lezgin, Maltese, Sicilian, Turkish_Jew, Tuscan
EEF	Native American	Adygei, Balkar, Iranian, Kumyk, North_Ossetian, Turkish
EEF	African	BedouinA, BedouinB†, Jordanian, Lebanese, Libyan_Jew, Moroccan_Jew, Palestinian, Saudi****, Syrian, Tunisian_Jew***, Yemenite_Jew***
EEF	South Asian	Armenian, Georgian****, Georgian_Jew*, Iranian_Jew***, Iraqi_Jew***

WHG = Loschbour or LaBraña; EEF = Stuttgart; ANE = MA1; Native American = Piapoco; African = Esan, Gambian, or Kgalagadi; South Asian = GujaratiC or Vishwabrahmin. Statistics are negative with $Z < -4$ unless otherwise noted: †(positive) or *, **, ***, ****, to indicate Z less than 0, -1, -2, and -3, respectively. The complete list of statistics can be found in Extended Data Table 1.

populations may themselves have been mixed. Indeed, the positive f_4 (Stuttgart, *Test*; Loschbour, Chimp) statistics obtained when *Test* is Near Eastern (Extended Data Table 1) imply that the EEF had some WHG-related ancestry, which was greater than 0% and as high as 45% (Supplementary Information section 13).

We used the ADMIXTUREGRAPH software^{8,15} to fit a model (a tree structure augmented by admixture events) to the data, exploring models relating the three ancient populations (Stuttgart, Loschbour, and MA1) to two eastern non-Africans (Onge and Karitiana) and sub-Saharan Africans (Mbuti). We found no models that fit the data with 0 or 1 admixture events, but did find a model that fit with 2 admixture events (Supplementary Information section 14). The successful model (Fig. 3) confirms the existence of MA1-related admixture in Native Americans³, but includes the novel inference that Stuttgart is partially ($44 \pm 10\%$) derived from a lineage that split before the separation of eastern non-Africans from the common ancestor of WHG and ANE. The existence of such basal Eurasian admixture into Stuttgart provides a simple explanation for our finding that diverse eastern non-African populations share significantly more alleles with ancient European and Upper Palaeolithic Siberian

hunter-gatherers than with Stuttgart (that is, f_4 (Eastern non-African, Chimp; Hunter-gatherer, Stuttgart) is significantly positive), but that hunter-gatherers appear to be equally related to most eastern groups (Supplementary Information section 14). We verified the robustness of the model by reanalysing the data using the unsupervised MixMapper⁷ (Supplementary Information section 15) and TreeMix²¹ software (Supplementary Information section 16), which both identified the same admixture events. The ANE–WHG split must have occurred $> 24,000$ years ago (as it must predate the age of MA1 (ref. 3)), and the WHG and Eastern non-African split must have occurred $> 40,000$ years ago (as it must predate the Tianyuan²² individual from China which clusters with Asians to the exclusion of Europeans). The basal Eurasian split must be even older, and might be related to early settlement of the Levant²³ or Arabia^{24,25} before the diversification of most Eurasians, or more recent gene flow from Africa²⁶. However, the basal Eurasian population shares much of the genetic drift common to non-African populations after their separation from Africans, and thus does not appear to represent gene flow between sub-Saharan Africans and the ancestors of non-Africans after the out-of-Africa bottleneck (Supplementary Information section 14).

Fitting present-day Europeans into the model, we find that few populations can be fit as two-way mixtures, but nearly all are compatible with three-way mixtures of ANE–EEF–WHG (Supplementary Information section 14). The mixture proportions from the fitted model (Fig. 4 and Extended Data Table 3) are encouragingly consistent with those obtained from a separate method that relates European populations to diverse outgroups using f_4 statistics, assuming only that MA1 is an unmixed descendent of ANE, Loschbour of WHG, and Stuttgart of EEF (Supplementary Information section 17). We infer that EEF ancestry in Europe today ranges from $\sim 30\%$ in the Baltic region to $\sim 90\%$ in the Mediterranean, consistent with patterns of identity-by-descent (IBD) sharing^{27,28} (Supplementary Information section 18) and shared haplotype analysis (chromosome painting)²⁹ (Supplementary Information section 19) in which Loschbour shares more segments with northern Europeans and Stuttgart with southern Europeans. Southern Europeans inherited their European hunter-gatherer ancestry mostly via EEF ancestors (Extended Data Fig. 6), whereas northern Europeans acquired up to 50% of WHG ancestry above and beyond what they received through their EEF ancestors. Europeans have a larger proportion of WHG than ANE ancestry in general. By contrast, in the Near East there is no detectable WHG ancestry, but up to $\sim 29\%$ ANE in the Caucasus (Supplementary Information section 14). A striking feature of these findings is that ANE ancestry is inferred to be present in nearly all Europeans today (with a maximum of $\sim 20\%$), but was absent in both farmers and hunter-gatherers from central and western Europe during the Neolithic transition. However, ANE ancestry was not completely absent from the larger European region at that time: we find that it was present in $\sim 8,000$ -years-old Scandinavian hunter-gatherers, as MA1 shares more alleles with Motala12 (SHG) than with Loschbour, and Motala12 fits as a mixture of 81% WHG and 19% ANE (Supplementary Information section 14).

Two sets of European populations are poor fits for the model. Sicilians, Maltese, and Ashkenazi Jews have EEF estimates of $> 100\%$, consistent with their having more Near Eastern ancestry than can be explained via

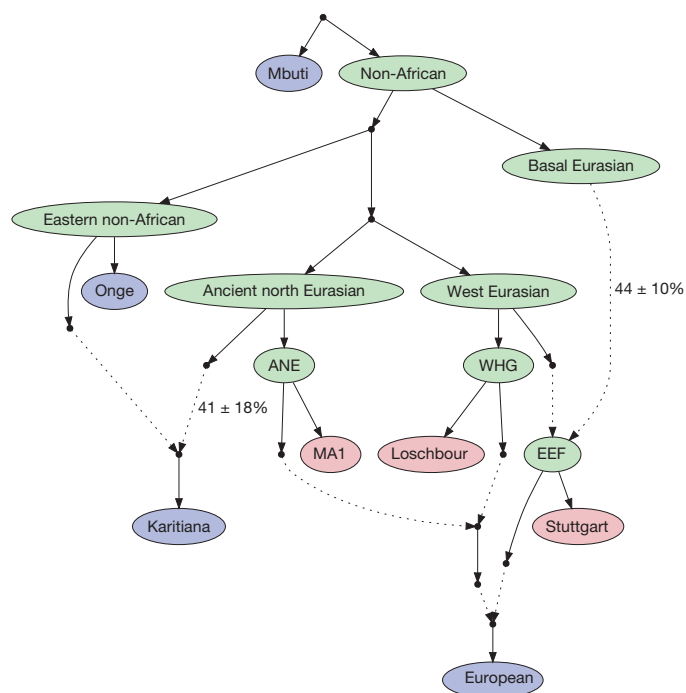


Figure 3 | Modelling the relationship of European to non-European populations. A three-way mixture model that is a fit to the data for many populations. Present-day samples are coloured in blue, ancient in red, and reconstructed ancestral populations in green. Solid lines represent descent without mixture, and dashed lines represent admixture. We print mixture proportions and one standard error for the two mixtures relating the highly divergent ancestral populations. (We do not print the estimate for the 'European' population as it varies depending on the population.)

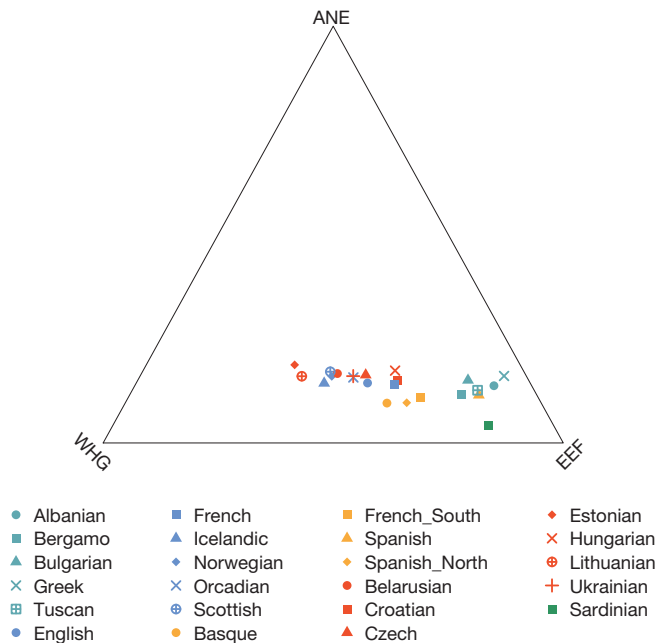


Figure 4 | Estimates of mixture proportions in present-day Europeans. Plot of the proportions of ancestry from each of three inferred ancestral populations (EEF, ANE and WHG).

EEF admixture (Supplementary Information section 17). They also cannot be jointly fit with other Europeans (Supplementary Information section 14), and they fall in the gap between European and Near Easterners in PCA (Fig. 2). Finns, Mordovians and Russians (from the north-west of Russia) also do not fit (Supplementary Information section 14; Extended Data Table 3) due to East Eurasian gene flow into the ancestors of these north-eastern European populations. These populations (and Chuvash and Saami) are more related to east Asians than can be explained by ANE admixture (Extended Data Fig. 7), probably reflecting a separate stream of Siberian gene flow into north-eastern Europe (Supplementary Information section 14).

Several questions will be important to address in future ancient DNA work. One question concerns where and when the Near Eastern farmers mixed with European hunter-gatherers to produce the EEF. A second question concerns how the ancestors of present-day Europeans first acquired their ANE ancestry. Discontinuity in central Europe during the late Neolithic (~4,500 years ago) associated with the appearance of mtDNA types absent in earlier farmers and hunter-gatherers³⁰ raises the possibility that ANE ancestry may have also appeared at this time. Finally, it will be important to study ancient genome sequences from the Near East to provide insights into the history of the basal Eurasians.

Online Content Methods, along with any additional Extended Data display items and Source Data, are available in the online version of the paper; references unique to these sections appear only in the online paper.

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Supplementary Information is available in the online version of the paper.

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Author Information The aligned sequences are available through the European Nucleotide Archive under accession number PRJEB6272. The fully public version of the Human Origins dataset can be found at (http://genetics.med.harvard.edu/reichlab/Reich_Lab/Datasets.html). The full version of the dataset (including additional samples) is available to researchers who send a signed letter to D.R. indicating that they will abide by specified usage conditions (Supplementary Information section 9). Reprints and permissions information is available at www.nature.com/reprints. The authors declare competing financial interests: details are available in the online version of the paper. Readers are welcome to comment on the online version of the paper. Correspondence and requests for materials should be addressed to D.R. (reich@genetics.med.harvard.edu) or J.Kr. (johannes.krause@uni-tuebingen.de).

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METHODS

Archaeological context, sampling and DNA extraction. The Loschbour sample stems from a male skeleton excavated in 1935 at the Loschbour rock shelter in Hefingen, Luxembourg. The skeleton was AMS radiocarbon dated to $7,205 \pm 50$ years before present (OxA-7738; 6,220–5,990 cal. BC)³¹. At the Palaeogenetics Laboratory in Mainz, material for DNA extraction was sampled from tooth 16 (an upper right M1 molar) after irradiation with ultraviolet light, surface removal, and pulverization in a mixer mill. DNA extraction took place in the palaeogenetics facilities in the Institute for Archaeological Sciences at the University of Tübingen. Three extracts were made in total, one from 80 mg of powder using an established silica based protocol³² and two additional extracts from 90 mg of powder each with a protocol optimized for the recovery of short DNA molecules³³.

The Stuttgart sample was taken from a female skeleton excavated in 1982 at the site Viesenhäuser Hof, Stuttgart-Mühlhausen, Germany. It was attributed to the Linearbandkeramik (5,500–4,800 BC) through associated pottery artefacts and the chronology was corroborated by radiocarbon dating of the stratigraphy³⁴. Both sampling and DNA extraction took place in the Institute for Archaeological Sciences at the University of Tübingen. Tooth 47 (a lower right M2 molar) was removed and material from the inner part was sampled with a sterile dentistry drill. An extract was made using 40 mg of bone powder³³.

The Motala individuals were recovered from the site of Kanaljorden in the town of Motala, Östergötland, Sweden, excavated between 2009 and 2013. The human remains at this site are represented by several adult skulls and one infant skeleton. All individuals are part of a ritual deposition at the bottom of a small lake. Direct radiocarbon dates on the remains range between $7,013 \pm 76$ and $6,701 \pm 64$ BP (6,361–5,516 cal. BC), corresponding to the late Middle Mesolithic of Scandinavia. Samples were taken from the teeth of the nine best preserved skulls, as well as a femur and tibia. Bone powder was removed from the inner parts of the teeth or bones with a sterile dentistry drill. DNA from 100 mg of bone powder was extracted³⁵ in the ancient DNA laboratory of the Archaeological Research Laboratory, Stockholm.

Library preparation. Illumina sequencing libraries were prepared using either double- or single-stranded library preparation protocols^{36,37} (Supplementary Information section 1). For high-coverage shotgun sequencing libraries, a DNA repair step with uracil DNA glycosylase (UDG) and endonuclease VIII (endo VIII) treatment was included in order to remove uracil residues³⁸. Size fractionation on a PAGE gel was also performed in order to remove longer DNA molecules that are more likely to be contaminants³⁷. Positive and blank controls were carried along during every step of library preparation.

Shotgun sequencing and read processing. All non-UDG-treated libraries were sequenced either on an Illumina Genome Analyzer Iix with $2 \times 76 + 7$ cycles for the Loschbour and Motala libraries, or on an Illumina MiSeq with $2 \times 150 + 8 + 8$ cycles for the Stuttgart library. We followed the manufacturer's protocol for multiplex sequencing. Raw overlapping forward and reverse reads were merged and filtered for quality³⁹ and mapped to the human reference genome (hg19/GRCh37/1000Genomes) using the Burrows–Wheeler Aligner (BWA)⁴⁰ (Supplementary Information section 2). For deeper sequencing, UDG-treated libraries of Loschbour were sequenced on 3 Illumina HiSeq 2000 lanes with 50-bp single-end reads, 8 Illumina HiSeq 2000 lanes of 100-bp paired-end reads and 8 Illumina HiSeq 2500 lanes of 101-bp paired-end reads. The UDG-treated library for Stuttgart was sequenced on 8 HiSeq 2000 lanes of 101-bp paired-end reads. The UDG-treated libraries for Motala were sequenced on 8 HiSeq 2000 lanes of 100-bp paired-end reads, with 4 lanes each for two pools (one of 3 individuals and one of 4 individuals). We also sequenced an additional 8 HiSeq 2000 lanes for Motala12, the Motala sample with the highest percentage of endogenous human DNA. For the Loschbour and Stuttgart high coverage individuals, diploid genotype calls were obtained using the Genome Analysis Toolkit (GATK)⁴¹.

Enrichment of mitochondrial DNA and sequencing. To test for DNA preservation and mtDNA contamination, non-UDG-treated libraries of Loschbour and all Motala samples were enriched for human mitochondrial DNA using a bead-based capture approach with present-day human DNA as bait⁴². UDG-treatment was omitted in order to allow characterization of damage patterns typical for ancient DNA⁴⁰. The captured libraries were sequenced on an Illumina Genome Analyzer Iix platform with $2 \times 76 + 7$ cycles and the resulting reads were merged and quality filtered³⁹. The sequences were mapped to the Reconstructed Sapiens Reference Sequence, RSRS⁴³, using a custom iterative mapping assembler, MIA⁴⁴ (Supplementary Information section 4).

Contamination estimates. We assessed if the sequences had the characteristics of authentic ancient DNA using four approaches. First, we searched for evidence of contamination by determining whether the sequences mapping to the mitochondrial genome were consistent with deriving from more than one individual^{44,45}. Second, for the high-coverage Loschbour and Stuttgart genomes, we used a maximum-likelihood-based estimate of autosomal contamination that uses variation at sites that are fixed in the 1000 Genomes data to estimate error, heterozygosity and contamination⁴⁶

simultaneously. Third, we estimated contamination based on the rate of polymorphic sites on the X chromosome of the male Loschbour individual⁴⁷ (Supplementary Information section 3) Fourth, we analysed non-UDG treated reads mapping to the RSRS to search for ancient DNA-typical damage patterns resulting in C→T changes at the 5'-end of the molecule¹⁰ (Supplementary Information section 3).

Phylogenetic analysis of the mitochondrial genomes. All nine complete mitochondrial genomes that fulfilled the criteria of authenticity were assigned to haplogroups using Haplofind⁴⁸. A Maximum Parsimony tree including present-day humans and previously published ancient mtDNA sequences was generated with MEGA⁴⁹. The effect of branch shortening due to a lower number of substitutions in ancient lineages was studied by calculating the nucleotide edit distance to the root for all haplogroup R sequences (Supplementary Information section 4).

Sex determination and Y-chromosome analysis. We assessed the sex of all sequenced individuals by using the ratio of (chrY) to (chrY + chrX) aligned reads⁵⁰. We downloaded a list of Y-chromosome SNPs curated by the International Society of Genetic Genealogy (ISOGG, <http://www.isogg.org>) v. 9.22 (accessed Feb. 18, 2014) and determined the state of the ancient individuals at positions where a single allele was observed and MAPQ ≥ 30 . We excluded C/G or A/T SNPs due to uncertainty about the polarity of the mutation in the database. The ancient individuals were assigned haplogroups based on their derived state (Supplementary Information section 5). We also used BEAST v1.7.51 (ref. 51) to assess the phylogenetic position of Loschbour using 623 males from around the world with 2,799 variant sites across 500 kb of non-recombining Y-chromosome sequence⁵² (Supplementary Information section 5).

Estimation of Neanderthal admixture. We estimate Neanderthal admixture in ancient individuals with the f_4 ratio or S statistic^{8,53,54} $\hat{\alpha} = f_4(\text{Altai, Denisova; Test, Yoruba}) / f_4(\text{Altai, Denisova; Vindija, Yoruba})$ which uses whole genome data from Altai, a high coverage (52×) Neanderthal genome sequence⁵⁵, Denisova, a high coverage sequence³⁷ from another archaic human population (31×), and Vindija, a low coverage (1.3×) Neanderthal genome from a mixture of three Neanderthal individuals from Vindija Cave in Croatia⁵³.

Inference of demographic history and inbreeding. We used the Pairwise Sequentially Markovian Coalescent (PSMC)⁵⁶ to infer the size of the ancestral population of Stuttgart and Loschbour as it changed over time. This analysis requires high quality diploid genotype calls and cannot be performed in the low-coverage Motala samples. To determine whether the low effective population size inferred for Loschbour is due to recent inbreeding, we plotted the time-to-most-recent common ancestor (TMRCA) along each of chromosomes 1–22 to detect runs of low TMRCA.

Analysis of segmental duplications and copy number variants. We built read-depth based copy number maps for the Loschbour, Stuttgart and Motala12 genomes in addition to the Denisova and Altai Neanderthal genome and 25 deeply sequenced modern genomes⁵⁵ (Supplementary Information section 7). We built these maps by aligning reads, subdivided into their non-overlapping 36-bp constituents, against the reference genome using the mrsFAST aligner⁵⁷, and renormalizing read-depth for local GC content. We estimated copy numbers in windows of 500 unmasked base pairs slid at 100-bp intervals across the genome. We called copy number variants using a scale space filter algorithm. We genotyped variants of interest and compared the genotypes to those from individuals sequenced as part of the 1000 Genomes Project⁵⁸.

Phenotypic inference. We inferred likely phenotypes (Supplementary Information section 8) by analysing DNA polymorphism data in the VCF format⁵⁹ using VCFtools (<http://vcftools.sourceforge.net>). For the Loschbour and Stuttgart individuals, we included data from sites not flagged as LowQuality, with genotype quality (GQ) of ≥ 30 , and SNP quality (QUAL) of ≥ 50 . For Motala12, which is of lower coverage, we included sites having at least $2 \times$ coverage and that passed visual inspection of the local alignment using samtools view (<http://samtools.sourceforge.net>)⁶⁰.

Human Origins data set. We report new data on 1,615 present-day humans from 147 worldwide populations genotyped on the Affymetrix Human Origins array, all of whom provided informed consent consistent with studies of population history. The Human Origins array consists of 14 panels of SNPs for which the ascertainment is well known^{8,61}. All population genetics analysis were carried out on a set of 594,924 autosomal SNPs, after restricting to sites that had > 90% completeness across 7 different batches of sequencing, and that had > 97.5% concordance with at least one of two subsets of samples for which whole-genome sequencing data were also available. We generated our full data set by merging the newly collected data with previously reported data, resulting in 2,722 individuals (208 populations), which we filtered to 2,345 individuals (203 populations) after removing outlier individuals or relatives based on visual inspection of PCA plots^{14,62} or model-based clustering analysis¹³. Whole genome amplified (WGA) individuals were not used in analysis, except for a Saami individual who we included because of the special interest of this population for northeastern European population history (Extended Data Fig. 7).

ADMIXTURE analysis. We merged all Human Origins genotype data with whole genome sequencing data from Loschbour, Stuttgart, MA1, Motala12, Motala_merge, and LaBraña. We then thinned the resulting data set to remove SNPs in linkage disequilibrium with PLINK 1.07 (ref. 63), using a window size of 200 SNPs advanced by 25 SNPs and an r^2 threshold of 0.4. We ran ADMIXTURE 1.23 (refs 13, 64) for 100 replicates with different starting random seeds, default fivefold cross-validation, and varying the number of ancestral populations K between 2 and 20. We assessed clustering quality using CLUMPP⁶⁵. We used the ADMIXTURE results to identify a set of 59 ‘west Eurasian’ (European/Near Eastern) populations based on values of a west Eurasian ancestral population at $K = 3$ (Supplementary Information section 9). We also identified 15 populations for use as ‘non-west Eurasian outgroups’ based on their having at least 10 individuals and no evidence of European or Near Eastern admixture at $K = 11$, the lowest K for which Near Eastern/European-maximized ancestral populations appeared consistently across all 100 replicates.

Principal components analysis. We used smartpca¹⁴ (version: 10210) from EIGENSOFT^{62,66} 5.0.1 to carry out principal components analysis (PCA) (Supplementary Information section 10). We performed PCA on a subset on individuals and then projected others using the lsqproject: YES option that gives an unbiased inference of the position of samples even in the presence of missing data (especially important for ancient DNA).

f_3 statistics. We use the f_3 statistic⁸ $f_3(\text{Test}; \text{Ref}_1, \text{Ref}_2) = \frac{1}{N} \sum_{i=1}^N (t_i - r_{1,i})(t_i - r_{2,i})$, where t_i , $r_{1,i}$ and $r_{2,i}$ are the allele frequencies for the i^{th} SNP in populations *Test*, *Ref*₁, *Ref*₂, respectively, to determine if there is evidence that the *Test* population is derived from admixture of populations related to *Ref*₁ and *Ref*₂ (Supplementary Information section 11). A significantly negative statistic provides unambiguous evidence of mixture in the *Test* population⁸. We allow *Ref*₁ and *Ref*₂ to be any Human Origins population with 4 or more individuals, or Loschbour, Stuttgart, MA1, Motala12, LaBraña. We assess significance of the f_3 statistics using a block jackknife⁶⁷ and a block size of 5 cM. We report significance as the number of standard errors by which the statistic differs from zero (Z -score). We also perform an analysis in which we constrain the reference populations to be (1) EEF (Stuttgart) and WHG (Loschbour or LaBraña), (2) EEF and a Near Eastern population, (3) EEF and ANE (MA1), or (4) any two present-day populations, and compute a Z_{diff} score between the lowest f_3 statistic observed in the data set, and the f_3 statistic observed for the specified pair.

f_4 statistics. We analyse f_4 statistics⁸ of the form $f_4(A, B; C, D) = \frac{1}{N} \sum_{i=1}^N (a_i - b_i)(c_i - d_i)$ to assess if populations A, B are consistent with forming a clade in an unrooted tree with respect to C, D. If they form a clade, the allele frequency differences between the two pairs should be uncorrelated and the statistic has an expected value of 0. We set the outgroup D to be a sub-Saharan African population or chimpanzee. We systematically tried all possible combinations of the ancient samples or 15 ‘non-west Eurasian outgroups’ identified by ADMIXTURE analysis as A, B, C to determine their genetic affinities (Supplementary Information section 14). Setting A as a present-day test population and B as either Stuttgart or BedouinB, we documented relatedness to C = (Loschbour or MA1) or C = (MA1 and Karitiana) or C = (MA1 or Han) (Extended Data Figs 4, 5 and 7). Setting C as a test population and (A, B) a pair from (Loschbour, Stuttgart, MA1) we documented differential relatedness to ancient populations (Extended Data Fig. 6). We computed D -statistics⁵³ using transversion polymorphisms in whole genome sequence data⁵⁵ to confirm robustness to ascertainment and ancient DNA damage (Extended Data Table 2).

Minimum number of source populations for Europeans. We used qpWave^{16,17} to study the minimum number of source populations for a designated set of Europeans (Supplementary Information section 12). We use f_4 statistics of the form $X(l, r) = f_4(l_0, l; r_0, r)$ where l_0, r_0 are arbitrarily chosen ‘base’ populations, and l, r are other populations from two sets L and R respectively. If $X(l, r)$ has rank r and there were n waves of immigration into R with no back-migration from R to L , then $r + 1 \leq n$. We set L to include Stuttgart, Loschbour, MA1, Onge, Karitiana, Mbuti and R to include 23 modern European populations who fit the model of Supplementary Information section 14 and had admixture proportions within the interval [0,1] for the method with minimal modelling assumptions (Supplementary Information section 17).

Admixture proportions for Stuttgart in the absence of a Near Eastern ancient genome. We used Loschbour and BedouinB as surrogates for ‘unknown hunter-gatherer’ and Near Eastern (NE) farmer populations that contributed to Stuttgart (Supplementary Information section 13). Ancient Near Eastern ancestry in Stuttgart is estimated by the f_4 ratio^{8,15} $f_4(\text{Outgroup}, X; \text{Loschbour}, \text{Stuttgart}) / f_4(\text{Outgroup}, X; \text{Loschbour}, \text{NE})$. A complication is that BedouinB is a mixture of NE and African ancestry. We therefore subtracted¹⁷ the effects of African ancestry using estimates of the BedouinB African admixture proportion from ADMIXTURE (Supplementary Information section 9) or ALDER⁶⁸.

Admixture graph modelling. We used ADMIXTUREGRAPH⁸ (version 3110) to model population relationships between Loschbour, Stuttgart, Onge, and Karitiana

using Mbuti as an African outgroup. We assessed model fit using a block jackknife of differences between estimated and fitted f statistics for the set of included populations (we expressed the fit as a Z score). We determined that a model failed if $|Z| > 3$ for at least one f statistic. A basic tree model failed and we manually amended the model to test all possible models with a single admixture event, which also failed. Further manual amendment to include 2 admixture events resulted in 8 successful models, only one of which could be amended to also fit MA1 as an additional constraint. We successfully fit both the Iceman and LaBraña into this model as simple clades and Motala12 as a two-way mixture. We also fit present-day west Eurasians as clades, two-way mixtures, or three-way mixtures in this basic model, achieving a successful fit for a larger number of European populations ($n = 26$) as three-way mixtures. We estimated the individual admixture proportions from the fitted model parameters. To test if fitted parameters for different populations are consistent with each other, we jointly fit all pairs of populations A and B by modifying ADMIXTUREGRAPH to add a large constant (10,000) to the variance term $f_3(A_0, A, B)$. By doing this, we can safely ignore recent gene flow within Europe that affects statistics that include both A and B.

Ancestry estimates from f_4 -ratios. We estimate EEF ancestry using the f_4 ratio^{8,15} $f_4(\text{Mbuti}, \text{Onge}; \text{Loschbour}, \text{European}) / f_4(\text{Mbuti}, \text{Onge}; \text{Loschbour}, \text{Stuttgart})$, which produces consistent results with ADMIXTUREGRAPH (Supplementary Information section 14). We use $f_4(\text{Stuttgart}, \text{Loschbour}; \text{Onge}, \text{MA1}) / f_4(\text{Mbuti}, \text{MA1}; \text{Onge}, \text{Loschbour})$ to estimate Basal Eurasian admixture into Stuttgart. We use $f_4(\text{Stuttgart}, \text{Loschbour}; \text{Onge}, \text{Karitiana}) / f_4(\text{Stuttgart}, \text{Loschbour}; \text{Onge}, \text{MA1})$ to estimate ANE mixture in Karitiana (Fig. 4). We use $f_4(\text{Test}, \text{Stuttgart}; \text{Karitiana}, \text{Onge}) / f_4(\text{MA1}, \text{Stuttgart}; \text{Karitiana}, \text{Onge})$ to lower bound ANE mixture into north Caucasian populations.

MixMapper analysis. We carried out MixMapper 2.0 (ref. 7) analysis, a semi-supervised admixture graph fitting technique. First, we infer a scaffold tree of populations without strong evidence of mixture relative to each other (Mbuti, Onge, Loschbour and MA1). We do not include European populations in the scaffold as all had significantly negative f_3 statistics indicating admixture. We then ran MixMapper to infer the relatedness of the other ancient and present-day samples, fitting them onto the scaffold as two- or three-way mixtures. The uncertainty in all parameter estimates is measured by block bootstrap resampling of the SNP set (100 replicates with 50 blocks).

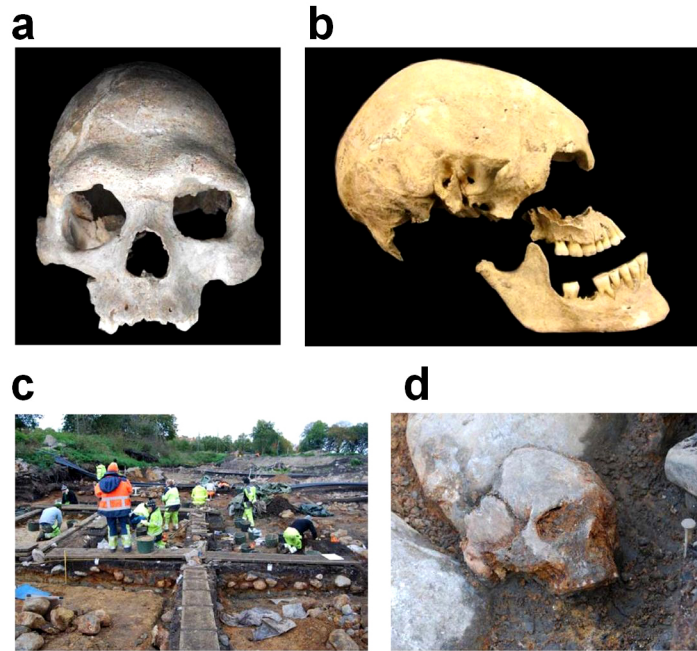
TreeMix analysis. We applied TreeMix²¹ to Loschbour, Stuttgart, Motala12, and MA1 (ref. 3), LaBraña² and the Iceman¹, along with the present-day samples of Karitiana, Onge and Mbuti. We restricted the analysis to 265,521 Human Origins array sites after excluding any SNPs where there were no-calls in any of the studied individuals. The tree was rooted with Mbuti and standard errors were estimated using blocks of 500 SNPs. We repeated the analysis on whole-genome sequence data, rooting with chimp and replacing Onge with Dai as we did not have Onge whole genome sequence data⁵⁵. We varied the number of migration events (m) between 0 and 5.

Inferring admixture proportions with minimal modelling assumptions. We devised a method to infer ancestry proportions from three ancestral populations (EEF, WHG, and ANE) without strong phylogenetic assumptions (Supplementary Information section 17). We rely on 15 ‘non-west Eurasian’ outgroups and study $f_4(\text{European}, \text{Stuttgart}; O_1, O_2)$ which is expected to equal $\alpha\beta f_4(\text{Loschbour}, \text{Stuttgart}; O_1, O_2) + \alpha(1-\beta) f_4(\text{MA1}, \text{Stuttgart}; O_1, O_2)$ if *European* has $1-\alpha$ ancestry from EEF and β , $1-\beta$ ancestry from WHG and ANE respectively. This defines a system of $\binom{15}{2} = 105$ equations with unknowns $\alpha\beta, \alpha(1-\beta)$, which we solve with least squares implemented in the function *lsfit* in R to obtain estimates of α and β . We repeated this computation 22 times dropping one chromosome at a time²⁰ to obtain block jackknife⁶⁷ estimates of the ancestry proportions and standard errors, with block size equal to the number of SNPs per chromosome. We assessed consistency of the inferred admixture proportions with those derived from the ADMIXTUREGRAPH model based on the number of standard errors between the two (Extended Data Table 1).

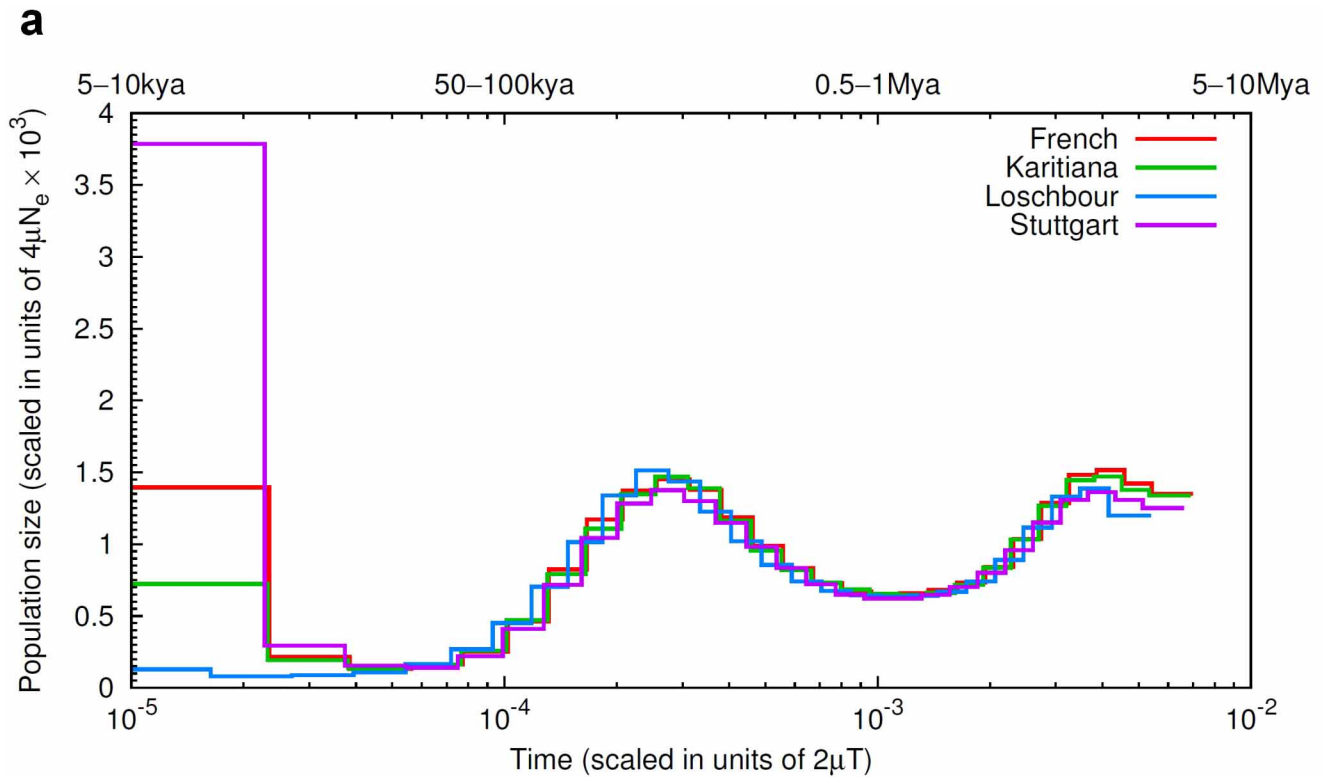
Haplotype-based analyses. We used RefinedIBD from BEAGLE 4²⁷ with the settings *ibdtrim* = 20 and *ibdwindow* = 25 to identify identity-by-descent (IBD) tracts: genomic segments or recently shared ancestry between Loschbour and Stuttgart and populations from the POPRES data set⁶⁹. We kept all IBD tracts spanning at least 0.5 centimorgans (cM) and with a LOD score > 3 (Supplementary Information section 18). We also used ChromoPainter²⁹ to study haplotype sharing between Loschbour and Stuttgart and present-day West Eurasian populations (SI19). We identified 495,357 SNPs that were complete in all individuals and phased the data using Beagle 4 (ref. 27) with parameters *phase-its* = 50 and *impute-its* = 10. We did not keep sites with missing data to avoid imputing modern alleles into the ancient individuals. We used both unlinked (-k 1000) and linked modes (estimating -n and -M by sampling 10% of individuals). We combined ChromoPainter output for chromosomes 1–22 using ChromoCombine²⁹. We carried out a PCA of the co-ancestry matrix using fineSTRUCTURE²⁹.

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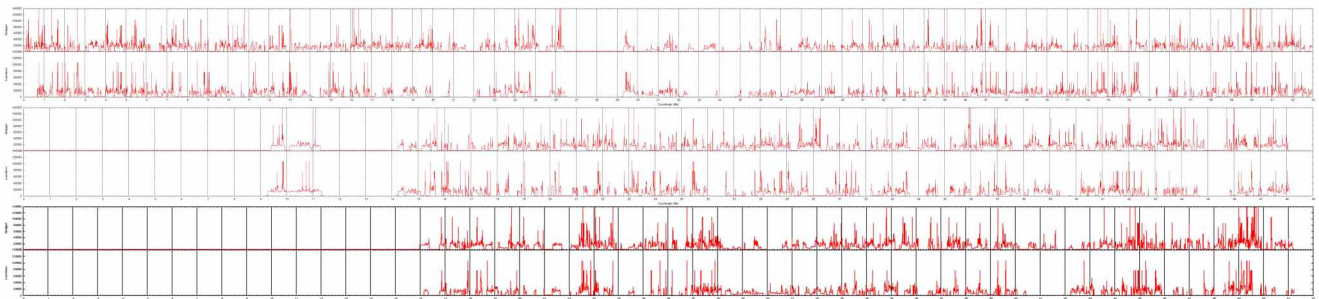
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Extended Data Figure 1 | Photographs of analysed ancient samples. a, Loschbour skull. b, Stuttgart skull, missing the lower right M2 we sampled. c, Excavation at Kanaljorden in Motala, Sweden. d, Motala 1 *in situ*.

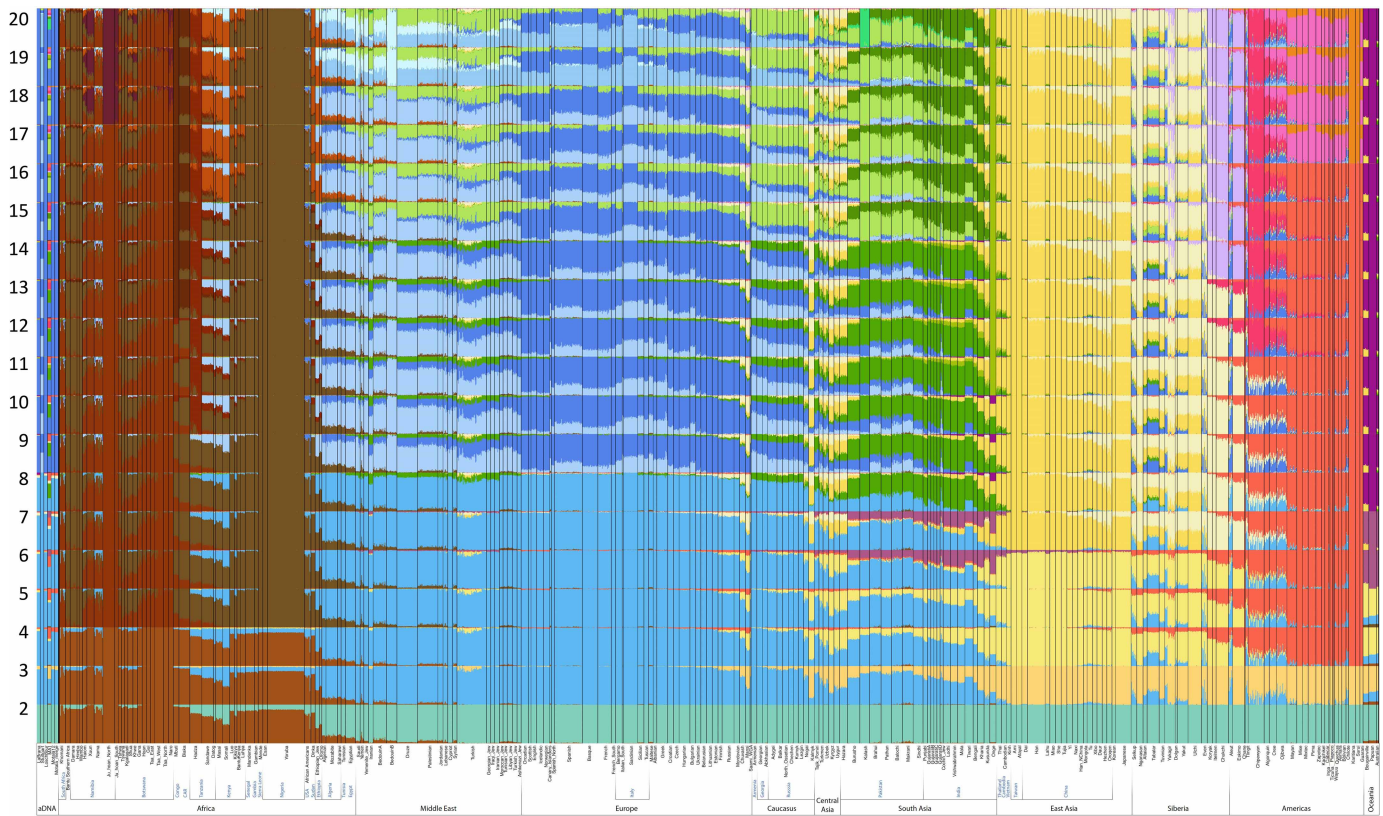


b

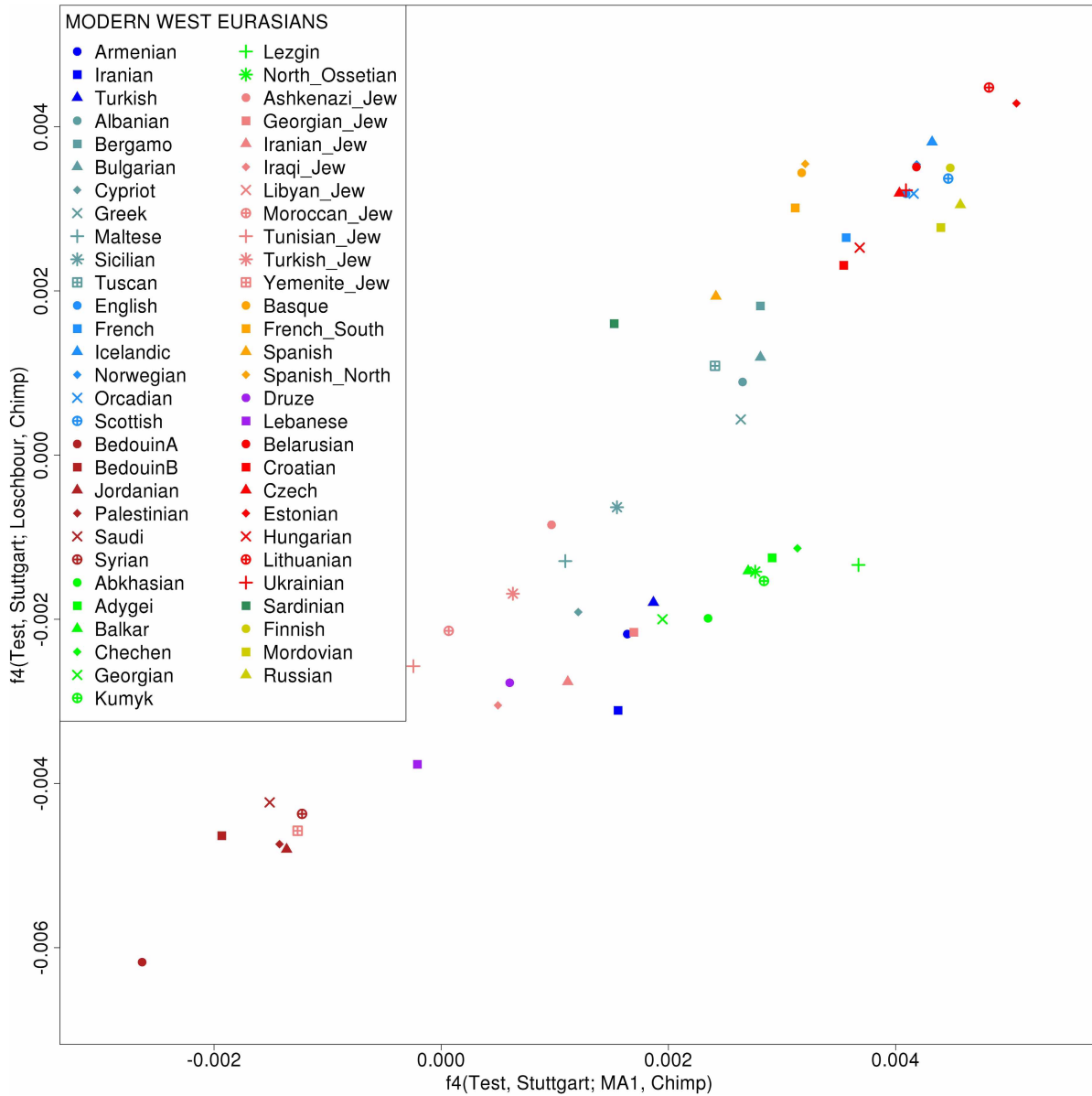


Extended Data Figure 2 | Pairwise sequential Markovian coalescent (PSMC) analysis. **a**, Inference of population size as a function of time, showing a very small recent population size over the most recent period in the ancestry

of Loschbour (at least the last 5–10 thousand years). **b**, Inferred time since the most recent common ancestor from the PSMC for chromosomes 20, 21, 22 (top to bottom); Stuttgart is plotted on top and Loschbour at bottom.

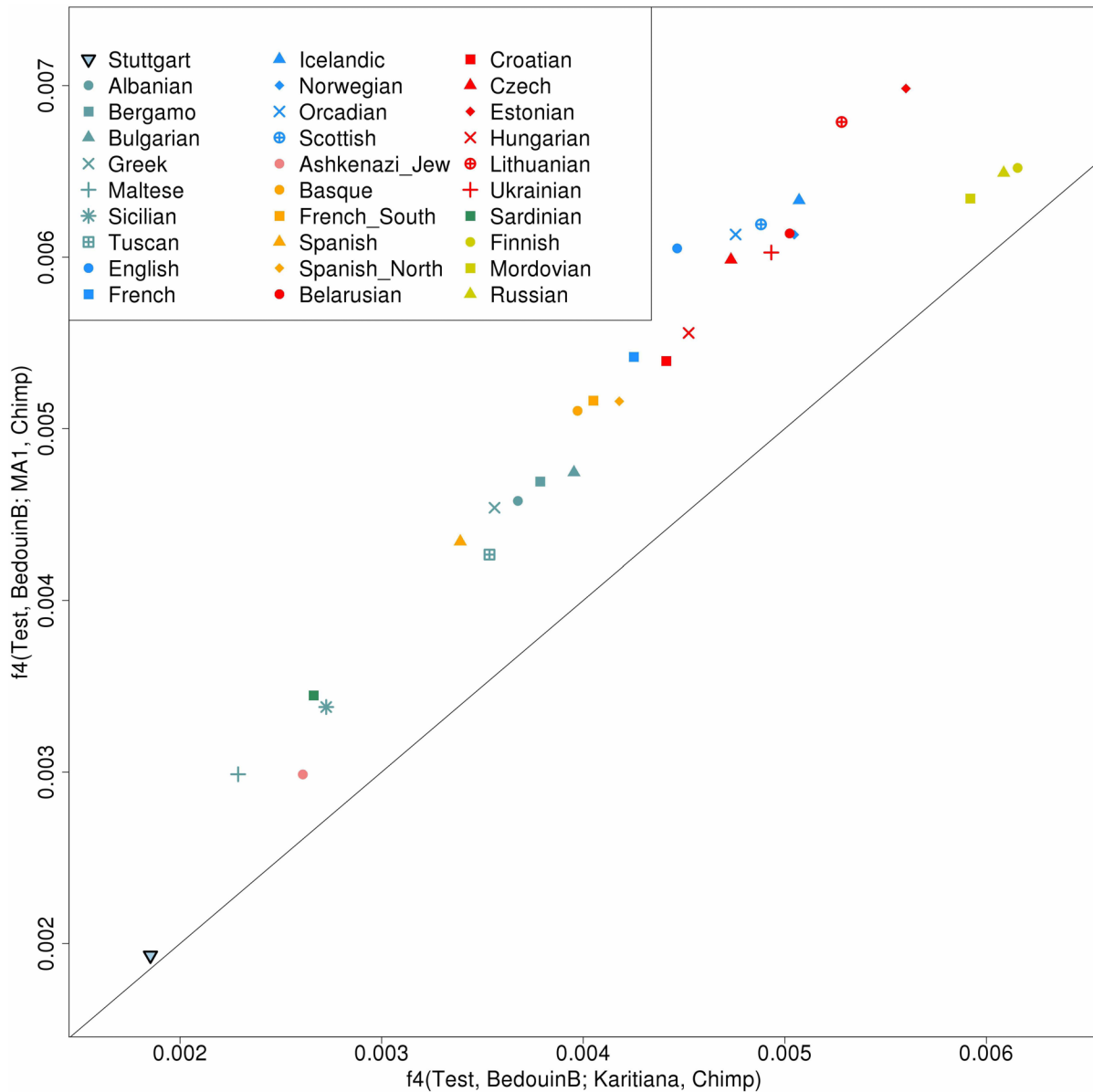


Extended Data Figure 3 | ADMIXTURE analysis ($K = 2$ to $K = 20$). Ancient samples (Loschbour, Stuttgart, Motala_merge, Motala12, MA1, and LaBraña) are on the left.



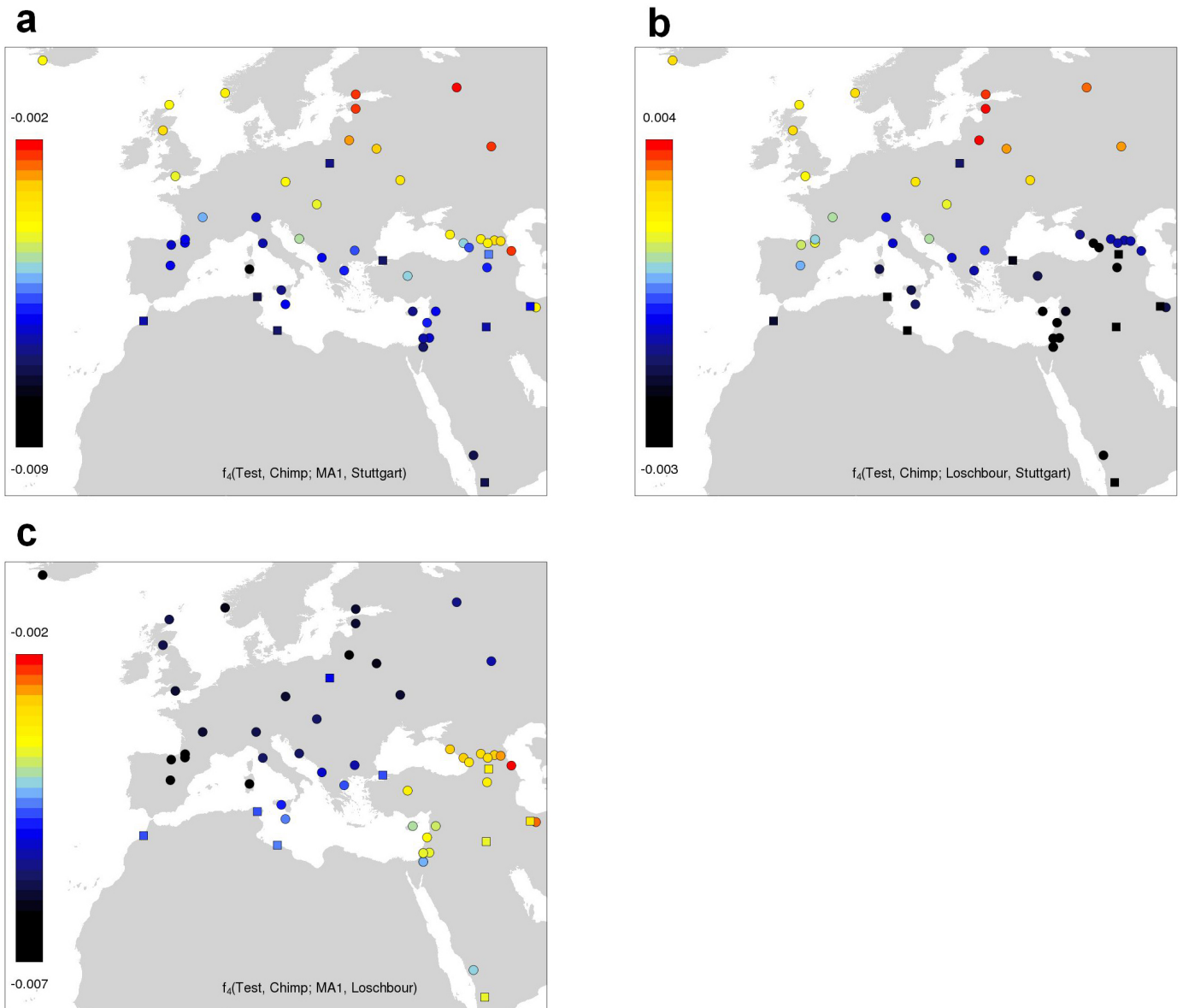
Extended Data Figure 4 | ANE ancestry is present in both Europe and the Near East but WHG ancestry is restricted to Europe, which cannot be due to a single admixture event. On the x axis we present the statistic $f_4(\text{Test}, \text{Stuttgart}; \text{MA1}, \text{Chimp})$, which measures where MA1 shares more alleles with a test population than with Stuttgart. It is positive for most European and Near Eastern populations, consistent with ANE (MA1-related) gene flow into

both regions. On the y axis we present the statistic $f_4(\text{Test}, \text{Stuttgart}; \text{Loschbour}, \text{Chimp})$, which measures whether Loschbour shares more alleles with a test sample than with Stuttgart. Only European populations show positive values of this statistic, providing evidence of WHG (Loschbour-related) admixture only in Europeans.



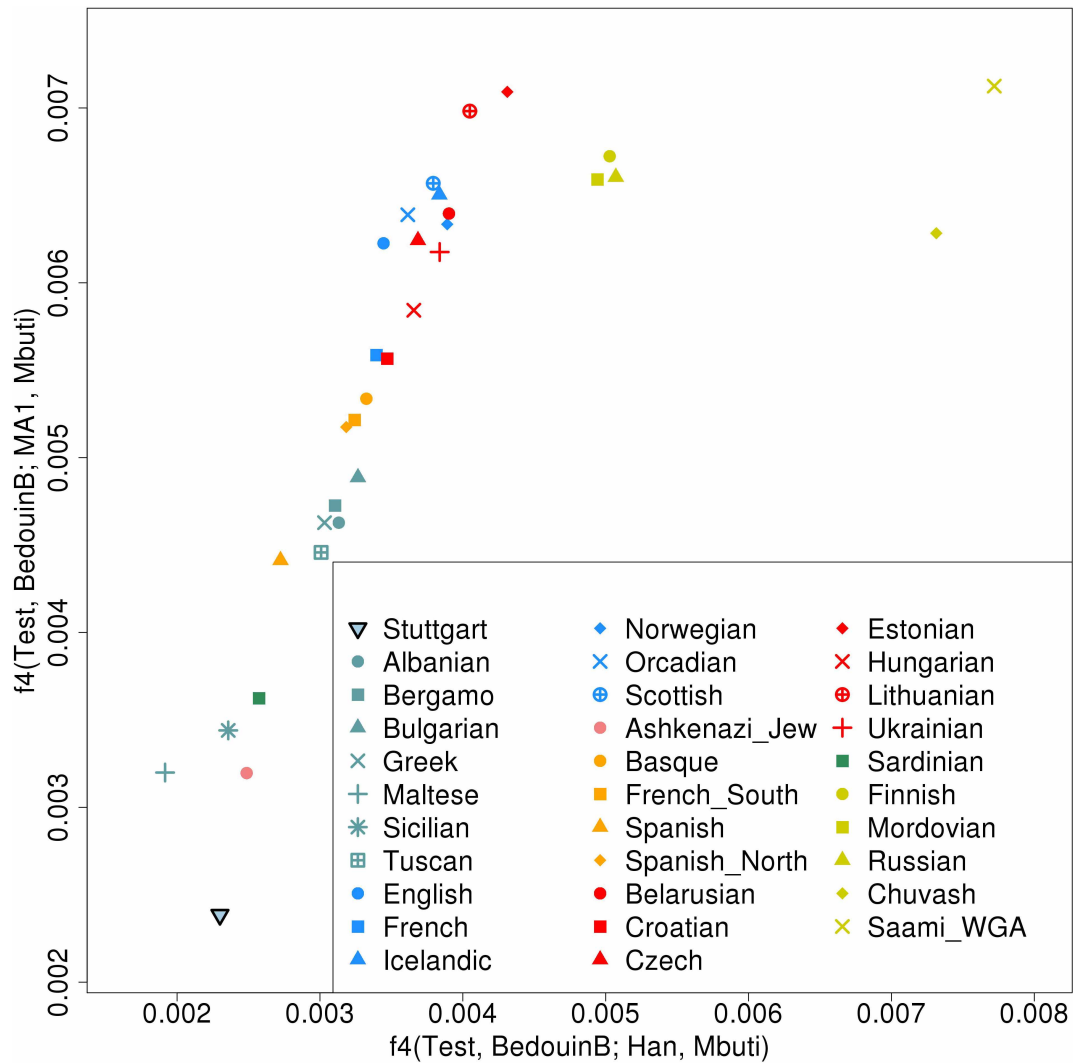
Extended Data Figure 5 | MA1 is the best surrogate for ANE for which we have data. Europeans share more alleles with MA1 than with Karitiana, as we see from the fact that in a plot of $f_4(\text{Test, BedouinB; MA1, Chimp})$ and

$f_4(\text{Test, BedouinB; Karitiana, Chimp})$, the European cline deviates in the direction of MA1, rather than Karitiana (the slope is > 1 and European populations are above the line indicating inequality of these two statistics).



Extended Data Figure 6 | The differential relatedness of west Eurasians to Stuttgart (EEF), Loschbour (WHG), and MA1 (ANE) cannot be explained by two-way mixture. We plot on a West Eurasian map the statistic $f_4(\text{Test, Chimp; } A_1, A_2)$, where A_1 and A_2 are a pair of the three ancient samples representing the three ancestral populations of Europe. **a**, In both Europe and the Near East/Caucasus, populations from the south have more relatedness to Stuttgart than those from the north where ANE influence is also important. **b**, Northern European populations share more alleles with Loschbour than with Stuttgart, as they have additional WHG ancestry beyond what was already

present in EEF. **c**, We observe a striking contrast between Europe west of the Caucasus and the Near East in degree of relatedness to WHG. In Europe, there is a much higher degree of allele sharing with Loschbour than with MA1, which we ascribe to the 60–80% WHG/(WHG + ANE) ratio in most Europeans that we report in Supplementary Information section 14. In contrast, the Near East has no appreciable WHG ancestry but some ANE ancestry, especially in the northern Caucasus. (Jewish populations are marked with a square in this figure to assist in interpretation as their ancestry is often anomalous for their geographic regions.)



Extended Data Figure 7 | Evidence for Siberian gene flow into far north-eastern Europe. Some north-eastern European populations (Chuvash, Finnish, Russian, Mordovian, Saami) share more alleles with Han Chinese

than with other Europeans who are arrayed in a cline from Stuttgart to Lithuanians/Estonians in a plot of $f_4(\text{Test, BedouinB; Han, Mbuti})$ against $f_4(\text{Test, BedouinB; MA1, Mbuti})$.

Extended Data Table 2 | Confirmation of key findings on transversions and on whole-genome sequence data

Interpretation	D(A, B; C, D) on Human Origins genotype data						D(A, B; C, D) on whole genome sequence data transversions							
	A	B	C	D	594,924 SNPs statistic	Z	110,817 transversions statistic	Z	A	B	C	D	statistic	Z
Stuttgart has Near Eastern ancestry	Stuttgart	Armenian	Loschbour	Chimp	0.0219	4.5	0.0189	2.9						
Europeans have more WHG-related ancestry than Stuttgart	Stuttgart	French	Loschbour	Chimp	-0.0266	-5.7	-0.031	-5.0	Stuttgart	French2	Loschbour	Chimp	-0.03	-4.7
		Lithuanian	Stuttgart	Loschbour	Chimp	0.0446	9.1	0.0477	7.2					
West Eurasians have more ANE-related ancestry than Stuttgart	French	Stuttgart	MA1	Chimp	0.0367	7.7	0.0386	5.5	French2	Stuttgart	MA1	Chimp	0.037	6.4
	Lezgin	Stuttgart	MA1	Chimp	0.0372	7.6	0.0409	5.6						
MA1 is a better surrogate of ANE ancestry than Karitiana	French	Chimp	MA1	Karitiana	0.0207	4.5	0.0214	2.8	French2	Chimp	MA1	Karitiana2	0.026	3.8
Eastern non-Africans closer to WHG/ANE/SHG than to EEF	Loschbour	Stuttgart	Onge	Chimp	0.0196	3.5	0.0202	2.5						
	Loschbour	Stuttgart	Papuan	Chimp	0.0142	2.6	0.0127	1.5	Loschbour	Stuttgart	Papuan2	Chimp	0.017	2.7
	Loschbour	Stuttgart	Dai	Chimp	0.0164	3.2	0.021	2.8	Loschbour	Stuttgart	Dai2	Chimp	0.018	2.9
	MA1	Stuttgart	Papuan	Chimp	0.0139	2.2	0.0103	1.0	MA1	Stuttgart	Papuan2	Chimp	0.018	2.8
	MA1	Stuttgart	Dai	Chimp	0.0174	3.0	0.016	1.7	MA1	Stuttgart	Dai2	Chimp	0.028	4.3
	Motala12	Stuttgart	Papuan	Chimp	0.0182	3.2	0.011	1.1	Motala12	Stuttgart	Papuan2	Chimp	0.023	3.7
	Motala12	Stuttgart	Dai	Chimp	0.0156	2.8	0.0149	1.6	Motala12	Stuttgart	Dai2	Chimp	0.02	3.2
	LaBrana	Stuttgart	Papuan	Chimp	0.0123	2.3	0.0101	1.1	LaBrana	Stuttgart	Papuan2	Chimp	0.02	3.2
LaBrana	Stuttgart	Dai	Chimp	0.0149	2.9	0.0228	2.5	LaBrana	Stuttgart	Dai2	Chimp	0.024	3.7	
Native Americans closer to ANE than to WHG	Karitiana	Chimp	MA1	Loschbour	0.0467	7.1	0.0467	4.4	Karitiana2	Chimp	MA1	Loschbour	0.052	7.1
West Eurasians closer to Native Americans than to other Eastern non-Africans	Stuttgart	Chimp	Karitiana	Papuan	0.0559	10.9	0.0474	6.6	Stuttgart	Chimp	Karitiana2	Papuan2	0.052	7.6
	Stuttgart	Chimp	Karitiana	Onge	0.0237	5.1	0.0179	2.6						
Ancient Eurasian hunter-gatherers equally related to Eastern non-Africans other than Native Americans	Loschbour	MA1	Dai	Chimp	-0.0015	-0.2	0.0016	0.2	Loschbour	MA1	Dai2	Chimp	-0.013	-1.9
	Loschbour	MA1	Papuan	Chimp	0.0002	0.0	0.0012	0.1	Loschbour	MA1	Papuan2	Chimp	-0.003	-0.4
	Loschbour	Motala12	Dai	Chimp	0.0024	0.4	0.009	0.9	Loschbour	Motala12	Dai2	Chimp	-0.002	-0.3
	Loschbour	Motala12	Papuan	Chimp	-0.0028	-0.4	0.0046	0.5	Loschbour	Motala12	Papuan2	Chimp	-0.004	-0.6
	MA1	Motala12	Dai	Chimp	0.0026	0.4	0.0047	0.4	MA1	Motala12	Dai2	Chimp	0.01	1.5
	MA1	Motala12	Papuan	Chimp	-0.0047	-0.7	-0.001	-0.1	MA1	Motala12	Papuan2	Chimp	-0.004	-0.5
LaBrana and Loschbour are a clade	LaBrana	Loschbour	Dai	Chimp	-0.0028	-0.5	0.0024	0.3	LaBrana	Loschbour	Dai2	Chimp	0.007	1.1
	LaBrana	Loschbour	Papuan	Chimp	-0.0031	-0.5	-0.0012	-0.1	LaBrana	Loschbour	Papuan2	Chimp	0.002	0.3
	LaBrana	Loschbour	MA1	Chimp	-0.006	-0.8	0.0101	0.7	LaBrana	Loschbour	MA1	Chimp	0.005	0.7
SHG closer to ANE than to WHG	Motala12	Loschbour	MA1	Chimp	0.0425	5.3	0.0353	2.6	Motala12	Loschbour	MA1	Chimp	0.042	5.9
	Motala12	LaBrana	MA1	Chimp	0.0465	5.8	0.0347	2.4	Motala12	LaBrana	MA1	Chimp	0.038	5.4
LaBrana and Loschbour equally related to Stuttgart	LaBrana	Loschbour	Stuttgart	Chimp	-0.0176	-2.6	-0.0106	-1.0	LaBrana	Loschbour	Stuttgart	Chimp	-0.012	-1.8

Extended Data Table 3 | Admixture proportions for European populations

	Full modeling of population relationships (individual fits)			Full modeling of population relationships (averaged fits)						Modeling of population relationships with minimal assumptions			Model-based (averaged) - Model with minimal assumptions (Z-score)		
	EEF	WHG	ANE	EEF		WHG		ANE		EEF	WHG	ANE	EEF	WHG	ANE
				Mean	Range	Mean	Range	Mean	Range						
Albanian	0.781	0.092	0.127	0.781	0.772-0.819	0.082	0.032-0.098	0.137	0.129-0.158	0.595 ± 0.112	0.353 ± 0.150	0.052 ± 0.049	1.658	-1.807	1.741
Ashkenazi_Jew	0.931	0	0.069							0.938 ± 0.146	-0.021 ± 0.185	0.083 ± 0.049			
Basque	0.593	0.293	0.114	0.569	0.527-0.616	0.335	0.255-0.392	0.096	0.076-0.129	0.569 ± 0.091	0.315 ± 0.124	0.115 ± 0.041	-0.001	0.165	-0.472
Belarusian	0.418	0.431	0.151	0.426	0.397-0.464	0.408	0.338-0.443	0.167	0.150-0.199	0.272 ± 0.094	0.554 ± 0.131	0.174 ± 0.047	1.637	-1.118	-0.158
Bergamo	0.715	0.177	0.108	0.721	0.704-0.793	0.163	0.061-0.189	0.117	0.104-0.147	0.644 ± 0.125	0.248 ± 0.170	0.108 ± 0.053	0.615	-0.503	0.162
Bulgarian	0.712	0.147	0.141	0.718	0.707-0.778	0.132	0.047-0.151	0.151	0.138-0.175	0.556 ± 0.110	0.328 ± 0.143	0.116 ± 0.043	1.469	-1.372	0.804
Croatian	0.561	0.293	0.145	0.564	0.548-0.586	0.285	0.242-0.310	0.151	0.137-0.172	0.453 ± 0.122	0.407 ± 0.159	0.140 ± 0.046	0.911	-0.768	0.238
Czech	0.495	0.338	0.167	0.489	0.460-0.531	0.348	0.273-0.382	0.163	0.145-0.196	0.402 ± 0.117	0.400 ± 0.162	0.198 ± 0.050	0.744	-0.322	-0.698
English	0.495	0.364	0.141	0.503	0.476-0.536	0.353	0.296-0.382	0.144	0.130-0.169	0.475 ± 0.091	0.357 ± 0.125	0.168 ± 0.043	0.304	-0.028	-0.561
Estonian	0.322	0.495	0.183	0.323	0.293-0.345	0.49	0.451-0.520	0.187	0.172-0.205	0.072 ± 0.121	0.778 ± 0.176	0.150 ± 0.064	2.070	-1.636	0.584
French	0.554	0.311	0.135	0.563	0.537-0.601	0.297	0.230-0.328	0.14	0.126-0.169	0.498 ± 0.097	0.359 ± 0.127	0.142 ± 0.039	0.672	-0.487	-0.060
French_South	0.675	0.195	0.13	0.636	0.589-0.738	0.256	0.111-0.323	0.108	0.088-0.151	0.636 ± 0.116	0.225 ± 0.165	0.140 ± 0.057	-0.003	0.189	-0.558
Greek	0.792	0.058	0.151	0.791	0.780-0.816	0.048	0.019-0.060	0.161	0.150-0.171	0.658 ± 0.098	0.255 ± 0.127	0.086 ± 0.039	1.357	-1.627	1.915
Hungarian	0.558	0.264	0.179	0.548	0.520-0.590	0.279	0.199-0.313	0.174	0.156-0.210	0.391 ± 0.109	0.454 ± 0.153	0.155 ± 0.050	1.437	-1.145	0.371
Icelandic	0.394	0.456	0.15	0.409	0.386-0.424	0.448	0.409-0.473	0.143	0.126-0.170	0.342 ± 0.102	0.476 ± 0.137	0.182 ± 0.045	0.654	-0.204	-0.861
Lithuanian	0.364	0.464	0.172	0.352	0.327-0.384	0.488	0.433-0.527	0.16	0.135-0.184	0.248 ± 0.117	0.548 ± 0.163	0.205 ± 0.052	0.886	-0.367	-0.864
Maltese	0.932	0	0.068							1.298 ± 0.185	-0.509 ± 0.248	0.211 ± 0.079			
Norwegian	0.411	0.428	0.161	0.417	0.388-0.438	0.423	0.383-0.450	0.16	0.140-0.181	0.273 ± 0.115	0.557 ± 0.161	0.170 ± 0.055	1.252	-0.831	-0.185
Orcadian	0.457	0.385	0.158	0.465	0.439-0.493	0.378	0.329-0.403	0.157	0.140-0.179	0.395 ± 0.088	0.437 ± 0.122	0.168 ± 0.041	0.798	-0.487	-0.264
Sardinian	0.817	0.175	0.008	0.818	0.791-0.874	0.141	0.058-0.182	0.041	0.026-0.068	0.883 ± 0.128	0.075 ± 0.166	0.042 ± 0.048	-0.510	0.400	-0.024
Scottish	0.39	0.428	0.182	0.408	0.387-0.424	0.421	0.384-0.448	0.171	0.149-0.201	0.286 ± 0.112	0.532 ± 0.156	0.182 ± 0.053	1.091	-0.712	-0.210
Sicilian	0.903	0	0.097							1.012 ± 0.149	-0.131 ± 0.199	0.119 ± 0.060			
Spanish	0.809	0.068	0.123	0.759	0.736-0.804	0.126	0.066-0.170	0.115	0.091-0.151	0.856 ± 0.126	-0.015 ± 0.165	0.160 ± 0.049	-0.769	0.855	-0.922
Spanish_North	0.713	0.125	0.163	0.612	0.561-0.660	0.292	0.214-0.365	0.096	0.072-0.126	0.581 ± 0.120	0.298 ± 0.158	0.121 ± 0.046	0.254	-0.038	-0.533
Tuscan	0.746	0.136	0.118	0.751	0.737-0.806	0.123	0.047-0.145	0.126	0.114-0.150	0.734 ± 0.118	0.153 ± 0.160	0.113 ± 0.054	0.141	-0.188	0.249
Ukrainian	0.462	0.387	0.151	0.463	0.445-0.491	0.376	0.322-0.399	0.16	0.148-0.187	0.259 ± 0.123	0.596 ± 0.173	0.145 ± 0.057	1.661	-1.269	0.269
Finnish										-0.299 ± 0.204	1.194 ± 0.296	0.105 ± 0.105			
Mordovian										-0.255 ± 0.173	1.151 ± 0.246	0.104 ± 0.090			
Russian										-0.303 ± 0.211	1.230 ± 0.301	0.072 ± 0.106			

The estimates from the model with minimal assumptions are from Supplementary Information section 17. The estimates from the full modelling are from Supplementary Information section 14 either by single population analysis or co-fitting population pairs and averaging over fits (these averages are the results plotted in Fig. 4). Populations that do not fit the models are not reported.