Genetic evidence for two founding populations of the Americas

Pontus Skoglund^{1,2}, Swapan Mallick^{1,2,3}, Maria Cátira Bortolini⁴, Niru Chennagiri^{1,2}, Tábita Hünemeier⁵, Maria Luiza Petzl-Erler⁶, Francisco Mauro Salzano⁴, Nick Patterson² & David Reich^{1,2,3}

Genetic studies have consistently indicated a single common origin of Native American groups from Central and South America¹⁻⁴. However, some morphological studies have suggested a more complex picture, whereby the northeast Asian affinities of present-day Native Americans contrast with a distinctive morphology seen in some of the earliest American skeletons, which share traits with present-day Australasians (indigenous groups in Australia, Melanesia, and island Southeast Asia)⁵⁻⁸. Here we analyse genome-wide data to show that some Amazonian Native Americans descend partly from a Native American founding population that carried ancestry more closely related to indigenous Australians, New Guineans and Andaman Islanders than to any present-day Eurasians or Native Americans. This signature is not present to the same extent, or at all, in present-day Northern and Central Americans or in a ~12,600year-old Clovis-associated genome, suggesting a more diverse set of founding populations of the Americas than previously accepted.

All Native American groups studied to date can trace all or much of their ancestry to a single ancestral population that probably migrated across the Bering land bridge from Asia more than 15,000 years ago² with some Northern American and Arctic groups also tracing other parts of their ancestry to more recent waves of migration^{2,9,10}. Ancient genomic evidence has shown that this so-called 'First American' ancestry is present in an individual associated with Clovis technology from North America dating to \sim 12,600 years ago³, and mitochondrial DNA has suggested that it was also present by 13,000-14,500 years ago^{11,12}. In contrast, some morphological analyses of early skeletons in the Americas have suggested that characteristics of some Pleistocene and early Holocene skeletons fall outside the variation of present-day Native Americans and instead fall within the variation of present-day indigenous Australians, Melanesians and so-called 'Negrito' groups from Southeast Asia (and some sub-Saharan African groups)7,13 This morphology has been hypothesized to reflect an initial 'Paleoamerican' pioneer population in the Americas, which according to some interpretations was largely replaced by populations with Northeast Asian affinities in the early Holocene, but may have persisted in some locations^{14,15}. However, morphological similarity can arise not only through shared descent but also through convergent evolution or phenotypic plasticity coupled with similar environments^{16,17}. Another limitation of morphological data is that it provides very few independent characters that can be analysed. Genome-wide data, with its hundreds of thousands of independent characters that evolve effectively neutrally, should be a statistically powerful and robust way to test whether a distinct lineage contributed to Native Americans.

Analysis of population history in the Americas is complicated by post-Columbian admixture from mainly European and African sources². We identified 63 individuals without discernable evidence of European or African ancestry in 21 Native American populations genotyped at \sim 600,000 single nucleotide polymorphisms (SNPs) on the Affymetrix Human Origins array 18,19 (Extended Data Fig. 1 and Supplementary Information section 1). We further restricted our studies to individuals from Central and South America that have the strongest evidence of deriving entirely from a homogeneous First American ancestral population². We computed all possible f_4 -statistics of the form $f_4(American_1, American_2; outgroup_1, outgroup_2)$, the product of the allele frequency differences between the two American groups and the two outgroups. We represented the Americans by a panel of 7 Central and South American groups, and the outgroups by 24 populations (4 from each of 6 worldwide regions). If the two Native American groups descend from a homogeneous ancestral population whose ancestors separated from the outgroups at earlier times, it follows that the difference in allele frequencies between Native American populations will have developed entirely after their separation from the outgroups, and so the correlation in allele frequency differences is expected to be zero. To evaluate whether all possible f_4 -statistics computed in this way are consistent with zero, correcting for multiple hypothesis testing due to the large number of statistics examined, we measured the empirical covariance of the matrix of f_4 -statistics using a block jackknife¹⁸, and performed a single Hotelling's T^2 test² for consistency with zero. We reject the null hypothesis at high significance $(P = 2 \times 10^{-7})$, suggesting that the analysed Native American populations do not all descend from a homogeneous ancestral population since separation from the outgroups (Extended Data Table 1 and Supplementary Information section 2). The coefficients for which non-American populations contribute the most to the signals separate Native Americans into a cline with two Amazonian groups (Suruí and Karitiana) on one extreme and Mesoamericans on the other (Extended Data Fig. 2). Among the outgroups, the most similar coefficients to Amazonian groups are found in Australasian populations: the Onge from the Andaman Islands in the Bay of Bengal (a so-called 'Negrito' group), New Guineans, Papuans and indigenous Australians (Supplementary Information section 2).

We extended our analysis to 197 non-American populations sampled worldwide¹⁸⁻²⁰. We computed D-statistics²¹ to test whether a randomly drawn derived allele from each worldwide population has an equal probability of matching a randomly drawn Mesoamerican or Amazonian chromosome at sites where these differ. This test takes as its null hypothesis the tree-like population history (Test population, (Mesoamericans, Amazonians)), and produces a positive D-statistic only in the case of excess affinity between the test population and Amazonians (negative values in the case of an excess affinity with Mesoamericans). Consistent with the signals observed when many populations are analysed together, we find that Andamanese Onge, Papuans, New Guineans, indigenous Australians and Mamanwa Negritos from the Philippines all share significantly more derived alleles with the Amazonians (4.6 > Z > 3.0 standard errors (s.e.) fromzero) (Extended Data Table 2). No population shares significantly more derived alleles with the Mesoamericans than with the Amazonians. We

¹Department of Genetics, Harvard Medical School, Boston, Massachusetts 02115, USA. ²Broad Institute of Harvard and MIT, Cambridge, Massachusetts 02142, USA. ³Howard Hughes Medical Institute, Harvard Medical School, Boston, Massachusetts 02115, USA. ⁴Departamento de Genética, Instituto de Biociências, Universidade Federal do Rio Grande do Sul, 91501-970 Porto Alegre, RS, Brazil. ⁵Departamento de Genética e Biologia Evolutiva, Universidade de São Paulo, 05508-090, SP, Brazil. ⁶Departamento de Genética, Universidade Federal do Paraná, 81531-980 Curitiba, PR, Brazil. find consistent results for this test not only for Onge, Papuans, New Guineans and indigenous Australians as representatives of Australasian populations, but also for different outgroups in place of chimpanzee: Africans, Europeans and East Asians (2.8 < Z < 4.8) (Supplementary Information section 3). In Fig. 1, we show a quantile–quantile plot of *D*-statistics contrasting the Mesoamerican Mixe and the Amazonian Suruı́, revealing Australasian populations as the only discernible outliers.

We replicated the significant evidence for affinity between Australasians and Amazonians using *D*-statistics computed on Illumina SNP array data² (as an alternative to the Affymetrix Human Origins SNP array data) (2.6 < Z < 3.0) and on high-coverage genome sequences from 3 Yoruba, 2 Suruí, 3 Mixe and 16 Papuans (18 of these genomes are reported for the first time here^{22,23}; Table 1) (Z = 4.3). In addition to the three independent molecular experiments that these data sets represent, we find consistent results for all different mutation classes in the high-coverage genomes (2.6 < Z < 4.3), and different ascertainment schemes (for example, in polymorphisms discovered in Africans, New Guineans and East Asians) (Supplementary Information section 3) (1.1 < Z < 3.3 for panels with >20,000 SNPs). We also find consistent results for two differently genotyped subsets of Suruí individuals from a total of 24 individuals² (Table 1 and Extended Data Fig. 3a) (2.6 < Z < 3.6). Simulations (Supplementary Information

section 3) show that genotype and sequence errors cannot explain the magnitude of the observed signal (Extended Data Fig. 3b). Finally, we generated new data from 9 populations from present-day Brazil using the Affymetrix Human Origins array, including previously untested individuals from the Amazonian Suruí and Karitiana for which DNA was extracted from blood. These new samples replicate the signal, and furthermore show that the signal is also strong in the Xavante (1.3 < Z < 3.25), a population of the Brazilian Central Plateau which speaks a language of the Ge group that is different from the Tupi language group to which the languages of the Karitiana and Suruí both belong. We do not detect any excess affinity to Australasians in the ~12,600-year-old Clovis-associated Anzick individual from western Montana (Z = -0.6) (Supplementary Information section 3).

To test if the significant *D*-statistics have the patterns expected for a genuine admixture event, we stratified the high coverage genomes into deciles of '*B*-values'²⁴, which measures proximity to functionally important regions. Genuinely significant *D*-statistics are expected to be of larger magnitude closer to genes, as selection increases variability in fitness of haplotypes near functionally important regions, which in turn increases the genetic drift in these regions and the absolute magnitude of *D*-statistics²⁵²⁶, a prediction that we confirmed empirically (Extended Data Fig. 4). We computed *D*(Yoruba, Papuan; Mixe,

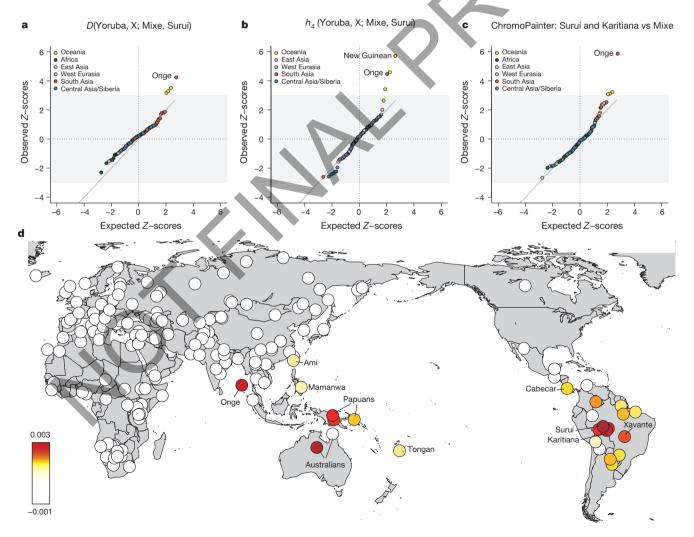


Figure 1 South Americans share ancestry with Australasian populations that is not seen in Mesoamericans or North Americans. a, Quantile–quantile plot of the *Z*-scores for the *D*-statistic symmetry test for whether Mixe and Suruí share an equal rate of derived alleles with a candidate non-American population *X*, compared to the expected ranked quantiles for the same number of normally distributed values. b, *Z*-scores for the h_4 -statistic. c, *Z*-scores for

the ChromoPainter statistic. **d**, Heatmap of ChromoPainter statistics. For non-Americans we display the symmetry statistic *S*(non-American; Mixe, Suruí and Karitiana) for donating as many haplotypes to Mixe as to Suruí and Karitiana. For the Americas we plot *S*(Onge; Mixe, American) for receiving as many haplotypes from the Onge as do the Mixe.

LETTER RESEARCH

	Test statistic	Z-score	Informative loci
High-coverage genomes	0.0211	4.26	798,873
A/T SNPs	0.0169	2.63	60,538
A/G SNPs	0.0191	3.64	268,962
A/C SNPs	0.0208	3.49	67,210
G/T SNPs	0.0248	4.27	67,623
C/T SNPs	0.0220	4.24	270,133
C/G SNPs	0.0248	4.26	64,951
Illumina array Suruí samples from HGDP	0.0076	2.63	247,814
Illumina array Suruí samples not in HGDP	0.0081	3.02	249,941
Affymetrix Human Origins array (Suruí cell lines)	0.0099	3.63	318,544
Affymetrix Human Origins array (Suruí blood samples)	0.0072	2.57	313,349
h ₄ -statistic (Affymetrix Yoruba ascertainment)	0.0003	4.60	14,938
Chromosome painting symmetry test	0.0026	5.26	-

Note: except for the new h₄ statistics and chromosome painting symmetry tests which are explicitly noted, all statistics are *D*-statistics²¹. *Z*-scores were obtained by computing standard errors using a weighted block jackknife.

Suruí) separately for each bin, and found that it is of larger magnitude close to functionally important regions (Extended Data Fig. 4) (Z = -2.0 for the slope of a linear regression model), as expected for a real admixture event. A caveat is that when we formally combine the evidence from the genome-wide *D*-statistic and the correlation to the *B*-value, the significance (Z = 3.6 s.e. from 0) is not any greater than for the basic $D = 0.021 \pm 0.005$ statistic (Z = 4.2 s.e. from 0) because the two statistics co-vary. Nevertheless, the fact that the correlation with *B*-values is significant by itself and in the expected direction adds to the qualitative evidence for an admixture event.

Alternative approaches for testing for admixture involve detecting admixture linkage disequilibrium in a test population that is correlated to allele frequency differentiation between two populations that are related to the sources^{27,28}. We devised a statistic ' h_4 ' that is analogous to an f_4 -statistic, but instead of studying allele frequencies, it tests whether the linkage disequilibrium patterns of two populations are consistent with descending from a common ancestral population since separation from two outgroups. A classic statistic for measuring linkage disequilibrium in a population A is $H^A = p_{12}^A - p_1^A p_2^A$, which measures the extent to which a haplotype of two derived mutations occurring at frequency p_{12}^A is observed more or less frequently than would be expected from the individual frequencies of alleles 1 and 2 $(p_1^A \text{ and } p_2^A)$. Thus, we define $h_4(A, B; C, D)$ as the average of $(H^A - H^B)(H^C - H^D)$ across the genome, and view a deviation from zero as evidence against the unrooted tree ((A, B), (C, D)). We used loci ascertained as polymorphic in African Yoruba, which is effectively an outgroup to the other populations analysed here, to test h_4 (Yoruba, X; Mixe, Suruí) for all SNP pairs within 0.01 centimorgans (cM) and for a large set of worldwide non-African populations, and obtained normalized Z-scores by estimating the number of standard errors this quantity is from zero using a block jackknife. Although Z-scores computed for most of 120 non-American and non-Africans as population X conform to a normal distribution (Fig. 1b), we again found significant evidence of excess affinity of the Suruí to Australasian populations $(Z = 5.7, P = 10^{-8} \text{ for New Guineans}; Z = 4.6, P = 10^{-5} \text{ for Papuans}; Z = 4.4, P = 10^{5} \text{ for Andamanese}$. When we exclude the Australasians, we detect no evidence of correlation between Z-transformed h_4 - and f_4 -statistics for the remaining 114 populations (R = -0.026) suggesting that h_4 can provide evidence independent of allele frequency based statistics. Although h_4 can theoretically be biased by loss of polymorphism due to bottlenecks (Supplementary Information section 4), there is no evidence that this is a problem for our analysis as East Asian and Siberian populations with comparable loss of polymorphism do not show an affinity to Amazonians by this statistic (Extended Data Fig. 5). In addition, there is a high degree of correlation between significant h_4 - and D-statistics in empirical data (Extended Data Fig. 5). Computing h_4 (Yoruba, Onge; Mixe, Suruí) over windows of increasingly large genetic distances reveals that it dissipates at approximately 0.2 cM. This is an order of magnitude smaller than linkage disequilibrium caused by admixture events at the ~4,000 year upper limit of previous methods¹⁸, but at a larger scale than the signal of admixture between Neanderthals and non-Africans 37,000-86,000 years ago²⁹ (Extended Data Fig. 5).

As a third population symmetry test, we applied a method for detecting shared haplotypes between individuals ('chromosome painting'30) to infer in each Native American individual which non-American chromosome segment each American chromosome segment shares the closest affinity to, using a set of 174 non-American populations as references. We then performed a symmetry test for a candidate population sharing more haplotypes with a given non-American population than the Mesoamerican Mixe do, performing a block jackknife across all chromosomes (weighting to correct for variation in chromosome length) to assess uncertainty. We find that the blood and cell line Suruí are significantly closer to the Onge than the Mixe are (Z = 5.3) (Fig. 1c), as are the blood and cell line Karitiana samples (Z = 4.2 to 5.0), the Xavante (Z=4.3), and the Piapoco and Guarani (Z>3) (Fig. 1d). In contrast, populations from west of the Andes or north of the Panama isthmus show no significant evidence of an affinity to the Onge (Z < 2). An exception to this is the Cabecar, who have previously been shown to be partially admixed from a source south of the Panama isthmus².

The geographic distribution of the shared genetic signal between South Americans and Australasians cannot be explained by post-Columbian African, European or Polynesian gene flow into Native American populations. If such gene flow produced signals strong enough to affect our statistics, our statistics would show their strongest deviations from zero for African, European or Polynesian populations, which is not observed. For example, a direct test is significant in showing that the Suruí-specific ancestry component is genetically closer to the Andamanese Onge than to Tongans from Polynesia (D = 0.0094, Z = 3.4).

To investigate models consistent with the data, we studied admixture graph models relating the ancestry of Native American groups to Han Chinese and Onge Andaman Islanders, incorporating a previously described admixture event into Native American ancestors from a lineage related to a ~24,000-year-old Upper Paleolithic individual from Mal'ta in Siberia⁴. We are unable to fit Amazonians as forming a clade with the Mesoamericans, or as having a different proportion of ancestry related to Mal'ta or present-day East Asians. Thus, our signal cannot be explained by lineages that have previously been documented as having contributed to Native American populations. However, we do find that a model where Amazonians receive ancestry from the lineage leading to the Andamanese fits the data in the sense that the predicted f_4 -statistics are all within two standard errors of statistics computed on the empirical data (Extended Data Figs 6 and 7 and Extended Data Table 3). These results do not imply that an unmixed population related anciently to Australasians migrated to the Americas. Although this is a formal possibility, an alternative model that we view as more plausible is that the 'Population Y' (after Ypykuéra, which means 'ancestor' in the Tupi language family spoken by the Suruí and Karitiana) that contributed Australasian-related

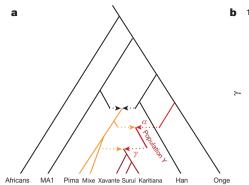
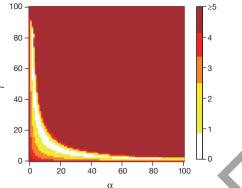


Figure 2 | A model of population history that can explain the excess affinity to Oceanians observed in Amazonian populations. a, We fit an admixture graph model where a population related to the Andamanese Onge contributed a fraction α of the ancestry of 'Population Y', which later contributed a fraction γ to the ancestry of Amazonian groups today

ancestry to Amazonians was already mixed with a lineage related to First Americans at the time it reached Amazonia. When we model such a scenario, we obtain a fit for models that specify 2–85% of the ancestry of the Suruí, Karitiana and Xavante as coming from Population Y (Fig. 2). These results show that quite a high fraction of Amazonian ancestry today might be derived from Population Y. At the same time, the results constrain the fraction of Amazonian ancestry that comes from an Australasian related population (via Population Y) to a much tighter range of 1–2% (Fig. 2).

We have shown that a Population Y that had ancestry from a lineage more closely related to present-day Australasians than to present-day East Asians and Siberians, likely contributed to the DNA of Native Americans from Amazonia and the Central Brazilian Plateau. This discovery is striking in light of interpretations of the morphology of some early Native American skeletons, which some authors have suggested have affinities to Australasian groups. The largest number of skeletons that have been described as having this craniofacial morphology and that date to younger than 10,000 years old have been found in Brazil⁶, the home of the Suruí, Karitiana and Xavante groups who show the strongest affinity to Australasians in genetic data. However, in the absence of DNA directly extracted from a skeleton with this morphology, our results are not sufficient to conclude that the Population Y we have reconstructed from the genetic data had this morphology.

An open question is when and how Population Y ancestry reached South America. There are several archaeological sites in the Americas that are contemporary to or earlier than Clovis sites. The fact that the one individual from a Clovis context who has yielded ancient DNA had entirely First American ancestry³ suggests the possibility that Population Y ancestry may be found in non-Clovis sites. Regardless of the archaeological associations, our results suggest that the genetic ancestry of Native Americans from Central and South America cannot be due to a single pulse of migration south of the Late Pleistocene ice sheets from a homogenous source population, and instead must reflect at least two streams of migration or alternatively a long drawn out period of gene flow from a structured Beringian or Northeast Asian source. The arrival of Population Y ancestry in the Americas must in any scenario have been ancient: while Population Y shows a distant genetic affinity to Andamanese, Australian and New Guinean populations, it is not particularly closely related to any of them, suggesting that the source of population Y in Eurasia no longer exists; furthermore, we detect no long-range admixture linkage disequilibrium in Amazonians as would be expected if the Population Y migration had occurred within the last few thousand years. Further insight into the population movements responsible for these findings should be



(the remainder of which is related to Mesoamerican Mixe). **b**, Twodimensional grid of combinations of the admixture proportions α and γ which are compatible with the data in terms of how many predicted f_4 -statistics deviate by $Z \ge 3.0$ from empirical values.

possible through genome-wide analysis of ancient remains from across the Americas.

Online Content Methods, along with any additional Extended Data display items and Source Data, are available in the online version of the paper; references unique to these sections appear only in the online paper.

Received 5 February; accepted 14 July 2015. Published online 21 July 2015.

- 1. Wang, S. et al. Genetic variation and population structure in Native Americans. PLoS Genet. **3**, e185 (2007).
- Reich, D. et al. Reconstructing Native American population history. Nature 488, 370–374 (2012).
- Rasmussen, M. et al. The genome of a Late Pleistocene human from a Clovis burial site in western Montana. Nature 506, 225–229 (2014).
- Raghavan, M. et al. Upper Palaeolithic Siberian genome reveals dual ancestry of Native Americans. Nature 505, 87–91 (2014).
- Neves, W. & Pucciarelli, H. The origins of the first Americans—an analysis based on the cranial morphology of early South American remains. *Am. J. Phys. Anthropol.* 81, 274 (1990).
- Neves, W. et al. Early Holocene human skeletal remains from Cerca Grande, Lagoa Santa, Central Brazil, and the origins of the first Americans. World Archaeol. 36, 479–501 (2004).
- Neves, W. A., Prous, A., González-José, R., Kipnis, R. & Powell, J. Early Holocene human skeletal remains from Santana do Riacho, Brazil: implications for the settlement of the New World. J. Hum. Evol. 45, 19–42 (2003).
- González-José, R. et al. Late Pleistocene/Holocene craniofacial morphology in Mesoamerican Paleoindians: implications for the peopling of the New World. Am. J. Phys. Anthropol. 128, 772–780 (2005).
- Rasmussen, M. et al. Ancient human genome sequence of an extinct Palaeo-Eskimo. Nature 463, 757–762 (2010).
- Raghavan, M. et al. The genetic prehistory of the New World Arctic. Science 345, (2014).
- Gilbert, M. T. P. et al. DNA from pre-Clovis human coprolites in Oregon, North America. Science 320, 786–789 (2008).
- Chatters, J. C. et al. Late Pleistocene human skeleton and mtDNA link Paleoamericans and modern Native Americans. Science 344, 750–754 (2014).
- Jantz, R. L. & Owsley, D. W. Variation among early North American crania. Am. J. Phys. Anthropol. **114**, 146–155 (2001).
- Neves, W. A., Hubbe, M. & Correal, G. Human skeletal remains from Sabana de Bogota, Colombia: a case of Paleoamerican morphology late survival in South America? *Am. J. Phys. Anthropol.* **133**, 1080–1098 (2007).
- González-José, R. et al. Craniometric evidence for Palaeoamerican survival in Baja California. Nature 425, 62–65 (2003).
- Sparks, C. S. & Jantz, R. L. A reassessment of human cranial plasticity: Boas revisited. Proc. Natl Acad. Sci. USA 99, 14636–14639 (2002).
- Relethford, J. H. Apportionment of global human genetic diversity based on craniometrics and skin color. Am. J. Phys. Anthropol. 118, 393–398 (2002).
- Patterson, N. et al. Ancient admixture in human history. Genetics 192, 1065–1093 (2012).
- Lazaridis, I. et al. Ancient human genomes suggest three ancestral populations for present-day Europeans. Nature 513, 409–413 (2014).
- Qin, P. & Stoneking, M. Denisovan ancestry in East Eurasian and Native American populations. *Mol. Biol. Evol.* (2015).
- Green, R. E. et al. A draft sequence of the Neandertal genome. Science 328, 710–722 (2010).
- Meyer, M. et al. Á high-coverage genome sequence from an Archaic Denisovan individual. Science 338, 222–226 (2012).

- Prüfer, K. et al. The complete genome sequence of a Neanderthal from the Altai Mountains. Nature 505, 43–49 (2014).
- McVicker, G., Gordon, D., Davis, C. & Green, P. Widespread genomic signatures of natural selection in hominid evolution. *PLoS Genet.* 5, e1000471 (2009).
- Gillespie, J. H. Genetic drift in an infinite population: the pseudohitchhiking model. Genetics 155, 909–919 (2000).
- Coop, G. et al. The role of geography in human adaptation. PLoS Genet. 5, e1000500 (2009).
- Moorjani, P. et al. The history of African gene flow into Southern Europeans, Levantines, and Jews. PLoS Genet. 7, e1001373 (2011).
- Hellenthal, G. et al. A genetic atlas of human admixture history. Science 343, 747–751 (2014).
- Sankararaman, Ś., Patterson, N., Li, H., Pääbo, S. & Reich, D. The date of interbreeding between Neandertals and modern humans. *PLoS Genet* 8, e1002947 (2012).
- Lawson, D. J., Hellenthal, G., Myers, S. & Falush, D. Inference of population structure using dense haplotype data. *PLoS Genet.* 8, e1002453 (2012).

Supplementary Information is available in the online version of the paper.

Acknowledgements We are grateful to the Native American volunteers who contributed the DNA samples used to generate the new data reported in this study and to the Fundação Nacional do Índio (FUNAI, Brazil) for logistical support in sample collection. We thank W. Klitz and C. Winkler for sharing samples for whole-genome sequencing. We thank L. Fehren-Schmitz, Q. Fu, G. Hellenthal, A. Kim, I. Lazaridis, M. Lipson, I. Mathieson, D. Meltzer, P. Moorjani and J. Pickrell for comments and A. Tandon for technical assistance. We thank T. Ferraz and R. Bisso-Machado for assistance with DNA extraction for the genotyping of Brazilian samples. We performed whole-genome sequencing as part of the Simons Genome Diversity Project. Genotyping of the Brazilian samples was performed at the Children's Hospital of Philadelphia and we particularly thank C. Hou for her support in this. M.C.B., T.H., M.L.P.-E. and F.M.S. were supported by Conselho Nacional do Desenvolvimento Científico e Tecnológico and Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (Brazil). P.S. was supported by the Wenner-Gren foundation and the Swedish Research Council (VR grant 2014-453). D.R. was supported by US National Science Foundation HOMINID grant BCS-1032255, US National Institutes of Health grant GM100233, Simons Foundation Grant 280376 and the Howard Hughes Medical Institute.

Author Contributions P.S. performed analyses. P.S., S.M., M.C.B., N.C., T.H., M.L.P.-E., F.M.S., N.P. and D.R. prepared datasets. P.S. and D.R. wrote the paper.

Author Information Genome sequence data is available from (https:// www.simonsfoundation.org/life-sciences/simons-genome-diversity-project-dataset/). New Affymetrix Human Origins array genotype data are available to researchers who send D.R. a signed letter agreeing to respect specific conditions (Supplementary Information section 1). Reprints and permissions information is available at www.nature.com/reprints. The authors declare no competing financial interests. Readers are welcome to comment on the online version of the paper. Correspondence and requests for materials should be addressed to P.S. (skoglund@genetics.med.harvard.edu) or D.R. (reich@genetics.med.harvard.edu)

METHODS

Data reporting. No statistical methods were used to predetermine sample size. The experiments were not randomized. The investigators were not blinded to allocation during experiments and outcome assessment.

New Affymetrix Human Origins genotypes. We generated new Affymetrix Human Origins array genotypes for 48 individuals from 9 populations from present-day Brazil (Apalaí, Arara, Guarani GN, Guarani KW, Karitiana, Suruí, Urubu Kaapor, Xavante and Zoró). Ethical approval for the sample collection was provided by the Brazilian National Ethics Commission (CONEP Resolution no. 123/98). CONEP also approved the oral consent procedure and the use of these samples in studies of population history and human evolution. Individual and/or tribal informed oral consents were obtained from participants who were not able to read or write. All sampling was coordinated by co-authors of this study (M.L.P.-E. and F.M.S.) and their collaborators, in a manner consistent with the Helsinki Declaration and Brazilian laws and regulations applicable at the time of sampling. Logistical support for the sample collection was provided by the Fundação Nacional do Índio (FUNAI). We curated the data in the same way that was reported in ref. 19 (Supplementary Information section 1). We computationally phased these data together with the previously published Affymetrix Human Origins SNP array data using SHAPEIT2 (ref. 31) with default parameters.

High-coverage genome sequencing and processing. We sent samples from 18 Papuan, Mixe, Suruí and Yoruba individuals to Illumina for deep-coverage sequencing using a non-PCR-based protocol as part of the Simons Genome Diversity Project. The sequence reads were mapped using the 'aln' algorithm of BWA (version 0.5.10)³² and genotypes were inferred using the unified genotyper from GATK³³ (version 2.5.2-gf57256b) These data are available from (https://www.simonsfoundation.org/life-sciences/simons-genome-diversity-project-dataset/). Briefly, sequence reads were stripped of adapters before alignment to the decoy version of the hg19 reference sequence (hs37d5). Read groups were added for identification and compatibility with GATK tools, before indel realignment and duplicate removal. The genotyping performed thereafter used a reference-free procedure that reduces reference bias. A specially developed filtering engine assigned filtering levels from 0 to 9 for each position in the genome. All population genetic analyses in this paper used the most stringent level of filtering (level 9).

Testing for more than one ancestral population of Central and South Americans. To investigate whether Central and South American populations are consistent with being derived from a single stream of ancestry, we applied the software qpWave² to ask the question whether the set of f_4 -statistics of the form $f_4(A = American_1, B = American_2; X = outgroup_1, Y = outgroup_1) =$ $(p_A - p_B)(p_X - p_Y)$ forms a matrix that is consistent with being of rank 0 (averaged over all SNPs, where p_A , p_B , p_X , and p_Y are the frequencies of an arbitrarily chosen allele in populations A, B, X and Y at each locus). If all these Native American populations descend from the same stream of migration into the Americas, then the f_4 -statistic relating each Native American population to each non-Native American population should be the same for all Native American populations, and in particular consistent with 0. Formally, to evaluate whether the f_4 -statistic matrix is consistent with being of rank 0, we compute a Hotelling's T^2 test that appropriately corrects for the correlation structure of the f_4 -statistics. We analysed 7 Native American populations each with at least 3 individuals with no detected post-Columbian admixture, and 4 populations from each of 6 worldwide regions as outgroups (Supplementary Information section 2).

D-statistic tests based on correlation in allele frequencies. To investigate whether a tree-like population history ((A, B), (X, Y)) is consistent with the data, for example, with A = chimpanzee, B = Onge, X = Mixe and Y = Suruí, we computed *D*-statistics^{18,21}

$$D(A,B;X,Y) = \frac{(p_A - p_B)(p_X - p_Y)}{(p_A + p_B - 2p_A p_B)(p_X + p_Y - 2p_X p_Y)}$$

over all SNPs, where p_A , p_B , p_X , and p_Y are the frequencies of an arbitrarily chosen allele in populations *A*, *B*, *X* and *Y* at each locus. We computed standard errors using a block jackknife weighted by the number of SNPs in each 5 cM (5 Mb in the case of high-coverage genome sequences) block in the genome^{34,35}. We report *Z*-scores as normalized Z = D/s.e. and we interpret statistics |Z| > 3 as being significantly different from 0. We only considered SNPs that were informative, in the sense that they are polymorphic both within (*A*,*B*) and (*X*,*Y*).

Correlation of signal to regions of functional importance. We divided the genome into 10 deciles of the '*B*-value' described in ref. 24, which integrates multiple genomic annotations into a single estimate of proximity to functional regions for each nucleotide in the genome. We then used linear regression to estimate the coefficient *a* of the function y = ax + c where x = B (the rank of the decile of *B*) and $y = D_B$ (*D* restricted to the particular decile of *B*). To compute

standard errors, we used a weighted block jackknife procedure where each 5 Mb block of the genome is dropped in turn and *a* is recomputed. The variability of *a* across each of these leave-one-out computations, weighting by the number of informative loci in each block, was what we used to estimate a standard error^{34,35} **h**₄-statistic tests based on correlation in linkage disequilibrium. We devised a linkage disequilibrium statistic that tests for symmetry in linkage disequilibrium between two proposed clades with a pair of populations in each. The statistic, *h*₄, is:

$$h_4 = \left(\left(p_{12}^A - p_1^A p_2^A \right) - \left(p_{12}^B - p_1^B p_2^B \right) \right) \times \left(\left(p_{12}^C - p_1^C p_2^C \right) - \left(p_{12}^D - p_1^D p_2^D \right) \right)$$

where *I* and *2* are arbitrarily chosen reference alleles at two different loci, respectively, and *A*, *B*, *C*, and *D* denote four different populations. Thus, p_{12}^A is the frequency of the *12* haplotype in population *A*, and p_1^A is the frequency of the *12* haplotype in population *A*, and p_1^A is the frequency of the *1* allele in population *A*. The quantity $p_{12}^A - p_1^A p_2^A$ thus measures the difference between the observed haplotype frequency and the expected haplotype frequency given the allele frequencies³⁶. The motivation for this statistic being informative about population history is that under a tree-like model ((*A*, *B*), (*C*, *D*)) with no gene flow, differences in linkage disequilibrium between populations *A* and *B* are not expected to correlate to differences in linkage disequilibrium between the statistic may be significantly positive or negative like f_4 and *D*-statistics¹⁸.

In practice, we computed this statistic for each polymorphic locus ('target locus') by identifying all other polymorphic loci 5' of the target locus at distance interval $d \pm w$ and computing the statistic for each pairing. We then averaged the statistic over all valid pairs of loci in the genome identified in this way. We computed standard errors using a block jackknife over contiguous 5 cM blocks in the genome, where SNP pairs that bridge the boundary of two blocks are assigned to the block in which the target locus is found. For the main analysis we computed h_4 -statistics of the form h_4 (Yoruba, X; Mixe, Surui) for all populations X genotyped using the Affymetrix Human Origins SNP array, and all pairs of SNPs within 0.01 cM of each other. We restricted the analysis to populations with at least 10 individuals. We also computed the h_4 -statistic for windows of 0.001 cM centred around different genetic distances for selected populations (Extended Data Fig. 5).

Chromosome-painting symmetry tests. We used SHAPEIT to phase 593,142 SNPs with the same set of individuals as described above, using all autosomal SNPs in the Affymetrix Human Origins array. We then 'painted' unadmixed Native American individuals using non-American populations, and excluded the Yukagir and the Chukchi since they have evidence of back-migration from the Americas. We ran ChromoPainter v2 using default parameters, painting each recipient individual separately, but using all donor populations as candidates to paint each recipient haplotype. To assess statistical uncertainty, we repeated this procedure for each recipient individual using 22 subsets of the data where for each of these subsets a different autosome had been dropped. We then used the results of these 22 block jackknife pseudo-replicates to obtain a weighted block jackknife estimate of the standard error for our test statistic (see below).

To test if the recipient populations copied equally from the donor populations, we computed the average 'chunk count' $C_{\text{R:D}}$ copied from a given donor population D in each recipient population R (averaged over individuals). We then computed a $S(R_1, R_2; D)$ statistic that quantifies the symmetry between two Native American populations in their copying from each donor:

$$S(D; R_2, R_1) = \frac{C_{R1:D} - C_{R2:D}}{C_{R1:D} + C_{R2:D}}$$

If two Native American populations, such as the Suruí and the Mixe, derive all of their ancestry from a single common origin, we expect that they would copy from the donor populations at an equal rate. We computed the standard error of this statistic using the 22 subsets of the data where each autosome had been dropped, weighted using the number of SNPs on each chromosome. We generated the world map in Fig. 1d by using the R maps package to plot the value of S(X; Mixe, Surui + Karitiana) for each non-American population X, and S(Onge; Mixe, Y) for each American population Y.

Admixture graph models of population relationships. We used ADMIXTURE-GRAPH¹⁸ to fit suggested phylogenies with admixture events to the data. We assessed goodness-of-fit by investigating all possible *f*-statistics predicted by the fitted model and assessing whether they differed significantly from the empirical data. We chose as a starting point the model relating Mbuti Africans, Andamanese Onge, MA1 and Karitiana fitted by a previous study¹⁹ where lineages related to MA1 and the Onge both contributed ancestry to the Karitiana. We added to this Han Chinese to represent a population that is phylogenetically more closely related to one of the ancestral populations of Native Americans than are the Onge (Extended Data Figs 6 and 7). We find that this model is inconsistent with the data, as the model predicts that Mixe and Suruí/Karitiana are equally related to Onge, and indeed we observe several statistics for which the Z-score for the



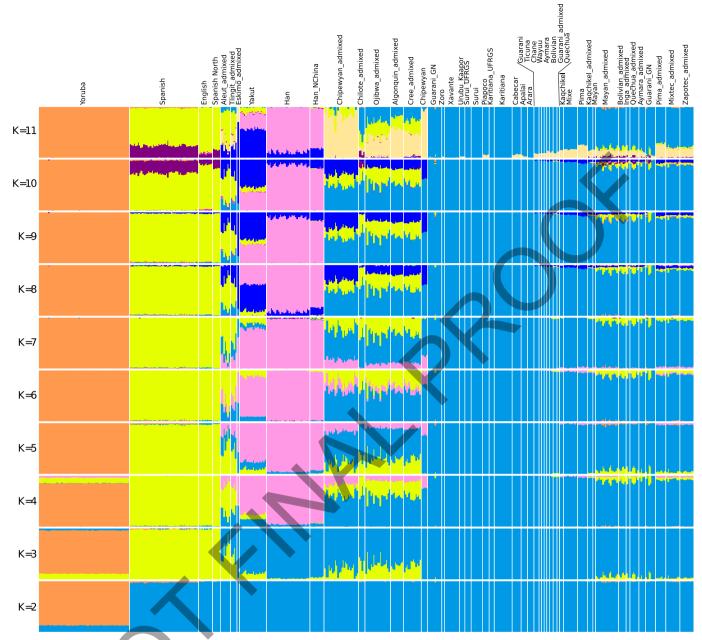
difference between the predicted and empirical statistics is |Z| > 3 (Extended Data Table 3). To account for this, we fitted a model in which the ancestors of Amazonians received admixture from a population related to the Onge (Extended Data Fig. 6), and found that this provides an excellent fit to the data, with no |Z|-score differences greater than 3. In contrast, alternative models of Han-related or MA1-related gene flow into the Americas are inconsistent with the data (Extended Data Fig. 6 and Extended Data Table 3).

Code availability. A python program for computing h_4 symmetry statistics and other population genetic statistics used in this paper is available at (https://github. com/pontussk/popstats).

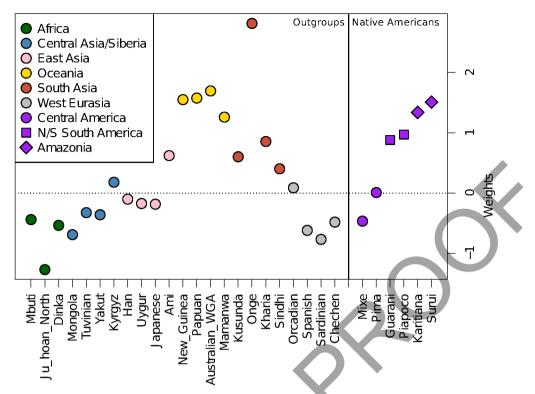
 Delaneau, O., Marchini, J. & Zagury, J.-F. A linear complexity phasing method for thousands of genomes. *Nature Methods* 9, 179–181 (2011).

- Li, H. & Durbin, R. Fast and accurate short read alignment with Burrows–Wheeler transform. *Bioinformatics* 25, 1754–1760 (2009).
- McKenna, A. et al. The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. Genome Res. 20, 1297–1303 (2010).
- Busing, F. M., Meijer, E. & Van Der Leeden, R. Delete-m jackknife for unequal m. Stat. Comput. 9, 3–8 (1999).
- Reich, D., Thangaraj, K., Patterson, N., Price, A. L. & Singh, L. Reconstructing Indian population history. *Nature* 461, 489–494 (2009).
- Robbins, R. B. Some applications of mathematics to breeding problems III. Genetics 3, 375–389 (1918).
- Becker, R. A. & Wilks, A. R. Maps in S. AT&T Bell Laboratories Statistics Research Report [93.2], (1993).
- Alexander, D. H., Novembre, J. & Lange, K. Fast model-based estimation of ancestry in unrelated individuals. *Genome Res.* 19, 1655–1664 (2009).

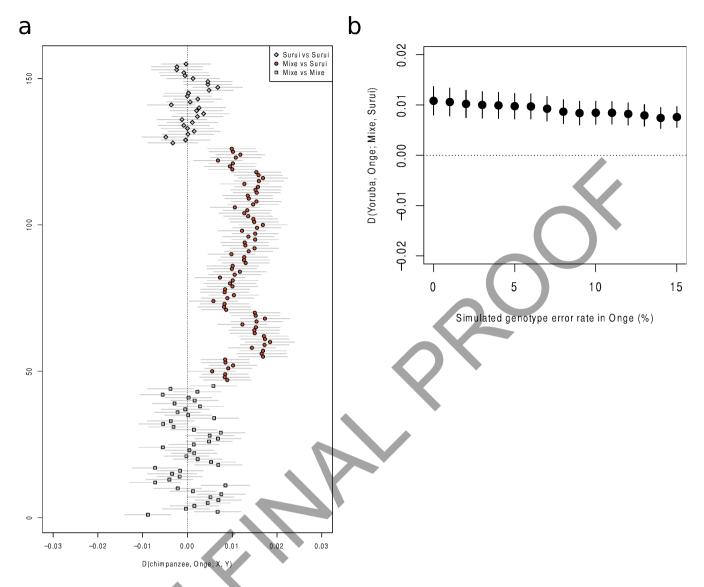
©2015 Macmillan Publishers Limited. All rights reserved



Extended Data Figure 1 Clustering analysis. ADMIXTURE³⁸ clustering analysis performed on the Affymetrix Human Origins data used in this study. To aid in visualization, we only show results for Native American samples and for selected samples from Eurasian populations.



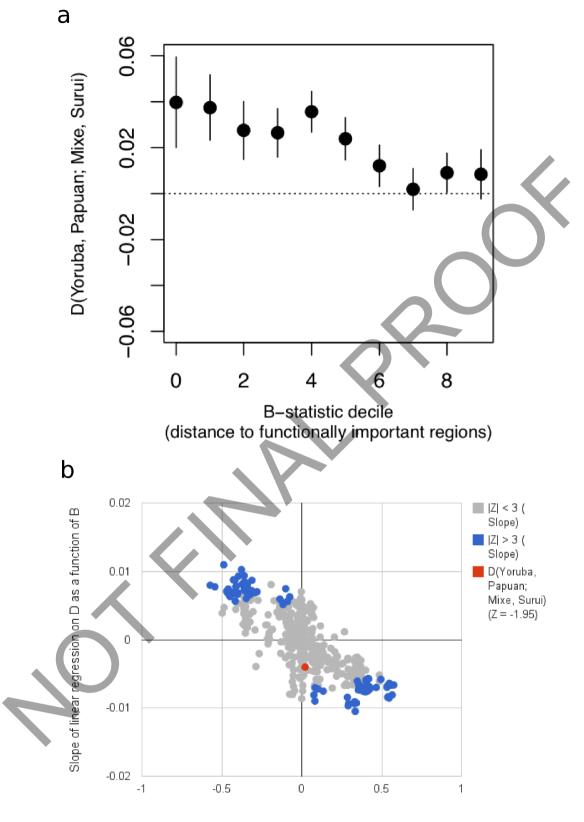
Extended Data Figure 2 | **qpWave coefficients.** Weights from qpWave for Native American populations and for non-American outgroup populations. No weights are given for Yoruba and Cabecar, as they are used in the computation.



Extended Data Figure 3 | **Excess allele sharing between the Surui and the Onge. a**, Tests for excess shared derived alleles with the Onge in all possible comparisons of 8 Suruí and 10 Mixe individuals. All Mixe–Suruí comparisons show a positive skew whereas all Mixe–Mixe and Suruí–Suruí comparisons

40

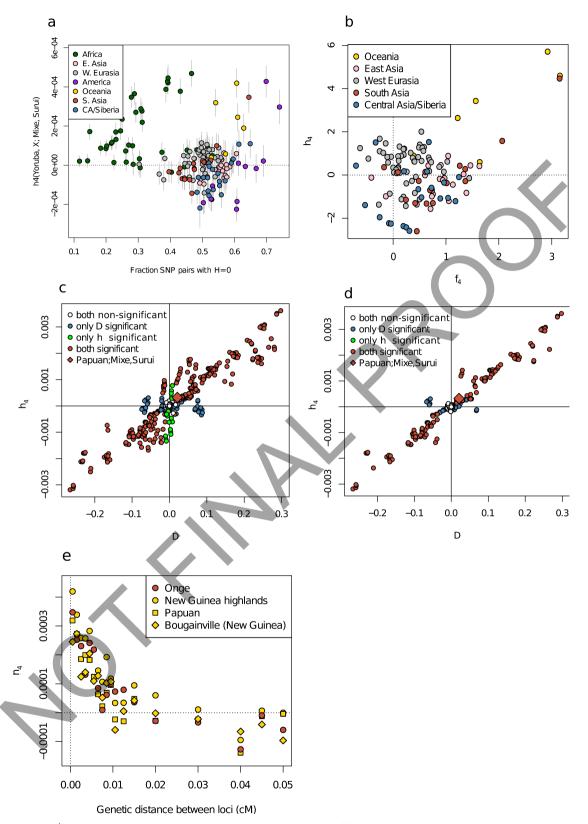
are consistent with 0. Lines correspond to one standard error in either direction. **b**, Random sequence or genotype errors cannot explain the affinity of the Amazonians to Australasians, as simulated increased errors in the Onge do not cause an increased affinity to Suruí.



Genome-wide D-statistic

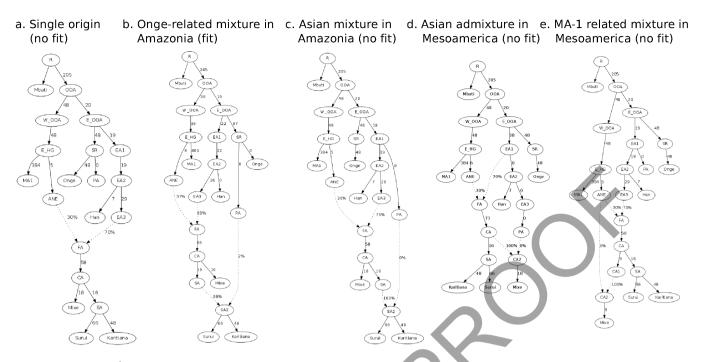
Extended Data Figure 4 | **Signals of admixture as a function of proximity to functional regions. a**, The affinity of 16 Papuan high-coverage genomes to 2 Amazonian Suruí high-coverage genomes as a function of proximity to regions of functional importance (measured by *B*-value). **b**, A total of 395 tests

of quartets D(Yoruba, X; Y, Z) shows that quartets with significantly positive slopes (|Z| > 3) also yield significant genome-wide D-statistics of the opposite sign. This suggests that signals of admixture are systematically stronger close to functionally important regions.



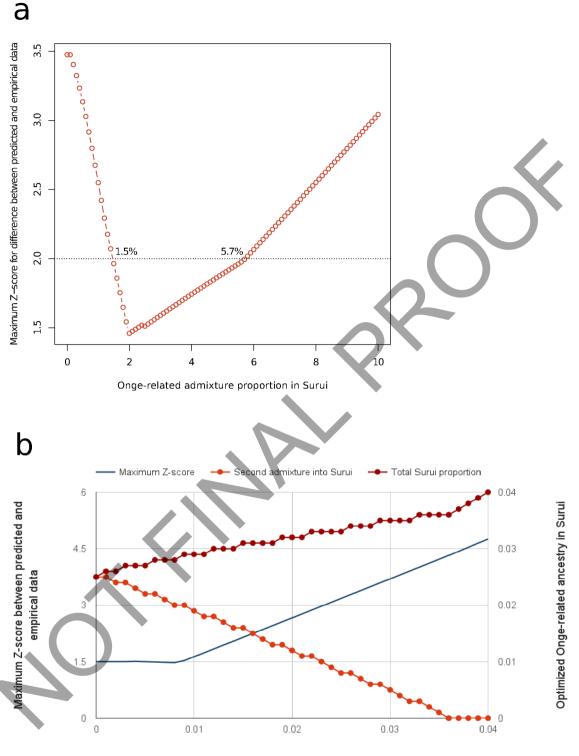
Extended Data Figure 5 | Linkage disequilibrium-based symmetry tests. a, h_4 (Yoruba, X; Mixe, Suruí) for SNP pairs within 0.01 cM of each other contrasted with the fraction of SNP pairs in linkage equilibrium in population X (H = 0). Error bars show ± 1 s.e. b, Scatterplot of *Z*-scores for the f_4 - and h_4 -statistics for the same quartets. For both these panels we only use populations with at least 6 samples. c, d, We computed *D*(Yoruba, *X*; *Y*, *Z*) and h_4 (Yoruba, *X*; *Y*, *Z*) for many combinations of populations as *X*, *Y* and *Z* using

phased Affymetrix Human Origins SNP array data ascertained in a Yoruba individual. Except for Africans who have ancestry from lineages that diverged before the Yoruba used for ascertainment and Oceanians (who have archaic Denisovan ancestry) we observe that $|Z| > 3 h_4$ -statistics are always associated with a significantly positive D for the same quartet. **e**, Correlation of the h_4 -statistic with the genetic distance separation of pairs of SNPs for h_4 (Yoruba, X; Mixe, Suruí).



Extended Data Figure 6 Admixture graphs for fitted population history models. a, An admixture graph where all of Mixe, Suruí and Karitiana are of 100% First American ancestry is rejected with 6 predicted *f*-statistics at least 3 standard errors from the empirically observed value. b, An admixture graph where the ancestors of Suruí and Karitiana receive 2% ancestry from a lineage related to the Onge is consistent with the data with no outliers. c, An admixture

graph where the distinct ancestry in Amazonians is more closely related to Han than to Onge produces 6 outliers. **d**, An admixture graph with no distinctive ancestry in Karitiana or Suruí but East Asian gene flow into the Mixe produces 7 outliers. **e**, An admixture graph with no distinctive ancestry in Karitiana or Suruí but MA1-related gene flow into the Mixe produces 6 outliers.



Assumed Onge-related ancestry in the Mixe

Extended Data Figure 7 | **Plausible range for the non-First American admixture proportion in Amazonians. a**, Range obtained assuming entirely First American ancestry in the Mixe. **b**, The maximum proportion of non-First American ancestry in the Mixe that is consistent with the data.

	<i>P</i> -value for this number of streams				
	1	2	3	4	
Full data	2.03E-07	0.09	0.58	0.92	
Outgroup region dropped	1				
Africa	1.67E-04	0.34	0.92	0.95	
C. Asia/Siberia	5.91E-07	0.11	0.6	0.89	
East Asia	4.46E-09	0.04	0.57	0.92	
South Asia	6.95E-05	0.1	0.4	0.82	
West Eurasia	1.41E-05	0.06	0.37	0.89	
Oceania	4.39E-05	0.43	0.88	0.97	
Native American populat	ion dropped				
Cabecar	1.13E-08	0.02	0.27	0.73	
Guarani	9.50E-07	0.27	0.76	0.99	
Karitiana	1.41E-06	0.1	0.61	0.86	
Mixe	8.32E-03	0.2	0.91	0.98	
Ріаросо	1.30E-04	0.55	0.93	0.98	
Pima	2.19E-05	0.31	0.78	0.97	
Surui	2.35E-06	0.11	0.56	0.84	
Africa + 1 other region					
Siberia	0.16	0.56	0.8	0.87	
East Asia	0.06	0.29	0.71	0.72	
South Asia	0.004	0.57	0.93	0.96	
West Eurasia	0.02	0.23	0.62	0.67	
Oceania	0.01	0.25	0.82	0.99	
Siberia + 1 other region					
Africa	0.16	0.56	0.8	0.87	
East Asia	0.41	0.91	0.99	1	
South Asia	0.03	0.82	0.91	0.9	
W. Eurasia	0.2	0.59	0.77	0.83	
Oceania	0.003	0.18	0.75	0.93	
	0.000	0.10	0.75		

Extended Data Table 1 | qpWave analysis provides evidence that Central and South American genetic variation is inconsistent with being derived from a single homogeneous population



Extended Data Table 2 | Top 20 D-statistics observed for D(chimpanzee, Old World population; Central Americans, Amazonians)

Rank	Population	D	SE	Ζ	Region 1	Region 2
1	Onge	0.0101	0.0022	4.60	India	South Asia
2	Papuan	0.0084	0.0022	3.82	Papua New Guinea	Oceania
3	New_Guinea	0.0082	0.0023	3.54	Papua New Guinea	Oceania
4	Australian_WGA	0.0074	0.0024	3.12	Australia (Arnhem Land)	Oceania
5	Mamanwa	0.0068	0.0020	3.40	Philippines (Negrito)	Oceania
6	Bougainville	0.0065	0.0023	2.85	Papua New Guinea	Oceania
7	Kharia	0.0059	0.0020	2.97	India	South Asia
8	Tongan	0.0058	0.0022	2.68	Tonga	Oceania
9	Bengali	0.0058	0.0019	3.00	Bangladesh	South Asia
10	Mala	0.0055	0.0019	2.93	India	South Asia
11	Ami	0.0052	0.0020	2.61	Taiwan	East Asia
12	Lodhi	0.0052	0.0019	2.72	India	South Asia
13	Sindhi	0.0051	0.0019	2.72	Pakistan	South Asia
14	Kusunda	0.0050	0.0020	2.56	Nepal	South Asia
15	Lahu	0.0050	0.0021	2.37	China	East Asia
16	Kinh	0.0049	0.0020	2.46	Vietnam	East Asia
17	Australian	0.0048	0.0025	1.96	Australia	Oceania
18	Balochi	0.0047	0.0019	2.55	Pakistan	South Asia
19	Thai	0.0047	0.0020	2.38	Thailand	East Asia
20	Semende	0.0045	0.0020	2.27	Indonesia (Sumatra)	Oceania

A	B	X	Y	Predicted f ₄		Z-score	
Single F	First Ame	erican ort	igin (Extende	ed Data Figure 6.	4)		
Mbuti	Onge	Mixe	Surui	0	0.003506	3.53	
Mbuti	Onge	Mixe	Karitiana	0	0.00315	3.43	
Onge	Mixe	Mixe	Surui	-0.018466	-0.021724	-3.06	
Onge	Mixe	Mixe	Karitiana	-0.018466	-0.021849	-3.22	
Onge	Han	Mixe	Surui	0	-0.002902	-3.65	
Onge	Han	Mixe	Karitiana	0	-0.00239	-3.27	
Onge-related ancestry in the Amazon (Extended Data Figure 6B)							
(No outliers)							
East Ast	ian admi			ca (Extended Date			
Mbuti	Onge	Mixe	Surui	0	0.003506	3.53	
Mbuti	Onge	Mixe	Karitiana	0	0.00315	3.43	
Onge	Mixe	Mixe	Surui	-0.018466	-0.021724	-3.06	
Onge	Mixe	Mixe	Karitiana	-0.018466	-0.021849	-3.22	
Onge	Han	Mixe	Surui	0	-0.002902	-3.65	
Onge	Han	Mixe	Karitiana	0	-0.00239	-3.27	
East Ast	ian admi	xture in (Central Ame	rica (Extended D	ata Figure 6D)		
Mbuti	Onge	Mixe	Surui	-0.000002	0.003506	3.53	
Mbuti	Onge	Mixe	Karitiana	-0.000002	0.00315	3.43	
Onge	Mixe	Mixe	Surui	-0.018466	-0.021724	-3.06	
Onge	Mixe	Mixe	Karitiana	-0.018466	-0.021849	-3.22	
Onge	Han	Mixe	Surui	-0.000004	-0.002902	-3.64	
Onge	Han	Mixe	Karitiana	-0.000004	-0.00239	-3.27	
Ancient	Siberiar	n (MA1) c	admixture in	Central America	(Extended Data I	Figure 6E	
Mbuti	Onge	Mixe	Surui	0	0.003506	3.53	
Mbuti	Onge	Mixe	Karitiana	0	0.00315	3.43	
Onge	Mixe	Mixe	Surui	-0.018470	-0.021724	-3.05	
Onge	Mixe	Mixe	Karitiana	-0.018470	-0.021849	-3.22	
Onge	Han	Mixe	Surui	0	-0.002902	-3.65	
Onge	Han	Mixe	Karitiana	0	-0.00239	-3.27	

Extended Data Table 3 | f_4 -statistics for which the statistic predicted by the fitted admixture graphs deviates by more than |Z| > 3 from the statistic computed on the empirical data

1. Data preparation

New Genotyping Data from 9 Brazilian Populations

We genotyped 48 individuals from 9 Brazilian Native American populations on the Affymetrix Human Origins array. All DNA was extracted from blood. We curated the data as in Lazaridis et al¹. Table S1.1 lists information on these new samples.

Sample ID	Sex	Population ID	Barion		Lat.	
Apalai147	U		Region Pará	Language Karib	-54.67	Long. -1.33
		Apalai				
Apalai185	M	Apalai	Pará	Karib	-54.67	-1.33
Apalai222	M	Apalai	Pará	Karib	-54.67	-1.33
Apalai228	M	Apalai	Pará	Karib	-54.67	-1.33
Arara1	M	Arara	Pará	Karib	-53.6	-3.9
Arara11	F	Arara	Pará	Karib	-53.6	-3.9
Arara23	М	Arara	Pará	Karib	-53.6	-3.9
Arara49	М	Arara	Pará	Karib	-53.6	-3.9
GuaraniGN5	F	Guarani_GN (Adm)	Mato Grosso do Sul	Tupi	-54.5	-23.33
GuaraniGN28	F	Guarani_GN (Adm)	Mato Grosso do Sul	Tupi	-54.5	-23.33
GuaraniGN405	F	Guarani_GN	Mato Grosso do Sul	Tupi	-54.5	-23.33
GuaraniGN841	F	Guarani_GN (Adm)	Mato Grosso do Sul	Tupi	-54.5	-23.33
GuaraniGN837	F	Guarani_GN (Adm)	Mato Grosso do Sul	Tupi	-54.5	-23.33
GuaraniGN845	F	Guarani_GN	Mato Grosso do Sul	Tupi	-54.5	-23.33
GuaraniGN852	F	Guarani_GN	Mato Grosso do Sul	Tupi	-54.5	-23.33
GuaraniKW203	F	Guarani_KW	Mato Grosso do Sul	Tupi	-55.2	-23.33
GuaraniKW220	F	Guarani_KW	Mato Grosso do Sul	Tupi	-55.2	-23.33
GuaraniKW223	F	Guarani_KW	Mato Grosso do Sul	Tupi	-55.2	-23.33
GuaraniKW224	F	Guarani_KW	Mato Grosso do Sul	Tupi	-55.2	-23.33
GuaraniKW230	F	Guarani_KW	Mato Grosso do Sul	Tupi	-55.2	-23.33
GuaraniKW644	F	Guarani_KW	Mato Grosso do Sul	Tupi	-55.2	-23.33
GuaraniKW645	F	Guarani_KW	Mato Grosso do Sul	Tupi	-55.2	-23.33
GuaraniKW646	М	Guarani_KW	Mato Grosso do Sul	Tupi	-55.2	-23.33
GuaraniKW650	F	Guarani_KW	Mato Grosso do Sul	Tupi	-55.2	-23.33
GuaraniKW626	F	Guarani_KW (Adm)	Mato Grosso do Sul	Tupi	-55.2	-23.33
Karitiana12	М	Karitiana	Rondônia	Tupi	-64.25	-9.33
Karitiana19	М	Karitiana	Rondônia	Tupi	-64.25	-9.33
Karitiana27	М	Karitiana	Rondônia	Tupi	-64.25	-9.33
Karitiana37	М	Karitiana	Rondônia	Tupi	-64.25	-9.33
Surui14	F	Surui	Rondônia	Tupi	-61.17	-10.33
Surui20	F	Surui	Rondônia	Tupi	-61.17	-10.33
Surui72	М	Surui	Rondônia	Tupi	-61.17	-10.33
Surui307	F	Surui	Rondônia	Tupi	-61.17	-10.33
UKaapor95	F	UrubuKaapor	Maranhão	Tupi	-45.22	-2.33
UKaapor150	M	UrubuKaapor	Maranhão	Tupi	-45.22	-2.33
UKaapor164	F	UrubuKaapor	Maranhão	Tupi	-45.22	-2.33
Xavante107	F	Xavante	Mato Grosso	Ge	-52.5	-14.33
Xavante214	U	Xavante	Mato Grosso	Ge	-52.5	-14.33
Xavante1004	M	Xavante	Mato Grosso	Ge	-52.5	-14.33
Xavante1104	F	Xavante	Mato Grosso	Ge	-52.5	-14.33
Xavante1105	F	Xavante	Mato Grosso	Ge	-52.5	-14.33
Xavante1513	F	Xavante	Mato Grosso	Ge	-52.5	-14.33
Xavante2302	M	Xavante	Mato Grosso	Ge	-52.5	-14.33
Xavante2302	M	Xavante	Mato Grosso	Ge	-52.5 -52.5	-14.33
Xavante401	F	Xavante	Mato Grosso	Ge Ge	-52.5 -52.5	
	F					-14.33
Xavante506		Xavante	Mato Grosso	Ge	-52.5	-14.33
Xavante606	F	Xavante	Mato Grosso	Ge	-52.5	-14.33
Zoro51	М	Zoro	Rondônia	Tupi	-60.33	-10.33

Table S1.1. New Human Origins Array genotypes for 48 Brazilian samples.

Note: Samples with "(Adm)" at the end of their population ID have population genetic evidence of European or African admixture.

The informed consent associated with these samples is not consistent with public posting of data. The data are available to researchers who send a PDF of a signed letter containing the text below to David Reich (reich@genetics.med.harvard.edu).

Box S1.1. Text that needs to be included in a letter to access the new data.

I affirm that

- (a) I will not distribute the data outside my collaboration
- (b) I will not post it publicly
- (c) I will make no attempt to connect the genetic data to personal identifiers
- (d) I will use the data only for studies of population history
- (e) I will not use the data for any commercial purposes

Identification of individuals without post-Columbian admixture

Previous genomic studies have identified substantial amounts of European and African ancestry due to post-Columbian admixture into Native American populations^{2,3}. However, since this admixture process is recent and ongoing there exists substantial intra-population variation in European and African ancestry in most admixed groups. To avoid the confounding factor of this admixture, we restricted analyses of Native Americans previously genotyped on the Affymetrix Human Origins array to individuals with <0.1% cluster membership in both the European-and African-modal components^{1,4} based on the *K*=3 ADMIXTURE analysis of Lazaridis et al¹. We excluded these individuals from subsequent analyses (Table S1.2).

PopulationIndividualsAdmixedPublished data	Not admixed 4 0 0 0 7 1 0 10 2 1 1 6 4
Chipewyan 30 26 Cree 13 13 Algonquin 9 9 Ojibwa 19 19 Pima 14 7 Mayan 18 17 Mixtec 10 0	0 0 7 1 0 10 2 1 6
Cree 13 13 Algonquin 9 9 Ojibwa 19 19 Pima 14 7 Mayan 18 17 Mixtec 10 10 Mixe 10 0	0 0 7 1 0 10 2 1 6
Algonquin 9 9 Ojibwa 19 19 Pima 14 7 Mayan 18 17 Mixtec 10 10 Mixe 10 0	0 0 7 1 0 10 2 1 6
Ojibwa 19 19 Pima 14 7 Mayan 18 17 Mixtec 10 10 Mixe 10 0	0 7 1 0 10 2 1 6
Pima 14 7 Mayan 18 17 Mixtec 10 10 Mixe 10 0	7 1 0 10 2 1 6
Mayan 18 17 Mixtec 10 10 Mixe 10 0	1 0 10 2 1 6
Mixtec 10 10 Mixe 10 0	0 10 2 1 6
Mixe 10 0	10 2 1 6
	2 1 6
	1 6
Kaqchikel 5 3	6
Wayuu 1 0	
Cabecar 6 0	4
Piapoco 4 0	
Inga 2 2	0
Ticuna 1 0	1
Karitiana 12 0	12
Surui 8 0	8
Quechua 7 6	1
Aymara 5 4	1
Chane 1 0	1
Guarani 5 2	3
Chilote 4 4	0
Bolivian 7 5	2
New data	
Apalai 4 0	4
Arara 4 0	4
Guarani_GN 6 4	2
Guarani_KW 11 1	10
Karitiana 4 0	4
Surui 4 0	4
Urubu_Kaapor 3 0	3
Xavante 11 0	11
Zoro 1 0	1
Total 240 133	107

 Table S1.2. Identification of admixed Native Americans.

For the newly genotyped Brazilians (Table S1.1) we used ADMIXTURE⁵ to infer cluster memberships. We co-analyzed the samples with other samples previously genotyped on the Human Origins array, after removing SNPs in high linkage disequilibrium using PLINK v1.07 (--indep-pairwise 200 25 0.4). We performed

clustering with ADMIXTURE for 2 to 12 clusters (*K*), using default parameters. We identified a Native American cluster at K=3 and excluded 4 Guarani_GN (Guarani-GN_5, Guarani-GN_28, Guarani-GN_837 and Guarani-GN_841), and 1 Guarani_KW (Guarani-KW_626) (Extended Data Figure 1).

Identification of unadmixed Dakelh

We analyzed data from 20 Athabascan-speaking Dakelh individuals from British Colombia⁶. These samples were genotyped on an Illumina SNP array which has only modest overlap with the Affymetrix Human Origins array, and so these samples had to be curated separately. To exclude individuals with evidence of recent European or East Asian admixture, we merged the samples with a diverse panel of individuals who had been previously genotyped using Illumina arrays⁷, and computed three statistics D(Yoruba, French; X, Karitiana), D(French, Han; Karitiana, X) and D(Yoruba, Han; X, Karitiana) for each Dalekh sample X in turn. We interpreted all statistics with |Z|>3 as providing significant evidence of post-Columbian admixture. Eleven samples had no evidence of mixture we restricted analyses to these (Table S1.3).

	D(Yoruba, Frenc	h; X, Karitiana)	D(French, Har	i; Karitiana, X)	D(Yoruba, Har	n; X, Karitiana)	
	D	Z	D	Z	D	Z	Status
athabaskSV6	0.003	1.10	-0.005	-1.66	-0.002	-0.81	
athabaskCN27	0.005	2.02	-0.014	-4.15	-0.009	-2.81	admixed
athabaskTL1	-0.004	-1.31	0.000	-0.12	-0.004	-1.26	
athabaskHD2	0.029	10.35	-0.072	-18.51	-0.044	-13.04	admixed
athabaskCN42	0.041	14.52	-0.105	-28.31	-0.065	-19.78	admixed
athabaskCA6	-0.005	-2.02	0.010	2.91	0.004	1.39	
athabaskCA12	-0.003	-1.15	0.007	2.36	0.004	1.23	
athabaskCA16	-0.001	-0.20	0.002	0.78	0.002	0.54	
athabaskCA24	0.003	1.03	-0.006	-2.06	-0.004	-1.24	
athabaskCA26	0.025	9.72	-0.064	-18.38	-0.039	-12.33	admixed
athabaskCA85	-0.003	-0.94	0.004	1.18	0.001	0.36	
athabaskCN9	0.013	4.24	-0.029	-8.06	-0.017	-5.36	admixed
athabaskCN40	0.000	0.03	-0.005	-1.61	-0.005	-1.56	
athabaskCA93	0.034	13.64	-0.099	-35.32	-0.065	-22.82	admixed
athabaskCA13	-0.006	-2.12	0.005	1.69	-0.001	-0.43	
athabaskSV3	-0.003	-1.11	0.008	2.64	0.005	1.57	
athabaskCN15	-0.002	-0.62	0.007	2.49	0.006	1.80	
athabaskCN36	0.011	3.57	-0.026	-7.71	-0.016	-4.98	admixed
athabaskHD4	0.048	18.63	-0.127	-37.98	-0.080	-25.46	admixed
athabaskHD3	0.034	12.66	-0.094	-25.09	-0.060	-18.28	admixed

Table S1.3. Identification of unadmixed Dakelh Athabascan-speakers.

Siberian groups with no evidence of recent back-from-America gene flow

A potential confounding factor for studies of migrations into the Americas is the use of far eastern Siberian populations who derive some ancestry from back-from-America gene flow². If the Native American ancestry in these Siberian populations is not symmetrically related to the Native American groups being studied, it has the potential to generate artifactual signals of distinct migrations into the Americas². In order to avoid using Siberian groups with such gene flow, we used ALDER⁸ to investigate whether there is evidence of mixture related to Native Americans in each Central Asian Siberian group in turn for which there is published Human Origins genotyping data. Specifically, we used the test for mixture implemented in ALDER, which tests for linkage disequilibrium (LD) that is correlated to the allele frequency differences between two populations that are proposed to be related to the admixing populations (here we use Han Chinese and Mixe Native Americans)⁸. We only consider populations with a 2-reference population Z-score of <3 for inclusion in the *qpWave* analysis in SI 2.

Test population	2-ref z-score	1-ref Z for Mixe	1-ref Z for Han	2-ref decay
Altaian	2.65	0	1.1	49 ± 14
Chukchi	2.1	3.49	7.83	28 ± 13
Eskimo	2.97	1.17	3.11	83 ± 18
Even	1.3	1.37	2.69	8 ± 6
Itelmen	1.38	0.39	0.17	84 ± 55
Kalmyk	2.5	2.17	3.99	42 ± 17
Koryak	2.53	1.7	1.63	51 ± 18
Kyrgyz	3.54	0	2.85	26 ± 7
Mansi	0.11	0.19	2.69	29 ± 262
Mongola	1.17	1.02	0	23 ± 20
Nganasan	0	1.16	3.29	n/a
Selkup	2.91	3.21	1.39	8 ± 3
Tajik_Pomiri	1.75	2.38	4.97	186 ± 66
Tubalar	3.55	2.17	9.7	33 ± 9
Turkmen	2.44	1.71	1.66	101 ± 41
Tuvinian	2.09	0	1.14	29 ± 13
Ulchi	1.35	2.09	4.03	167 ± 88
Uzbek	0.57	2.51	5.78	6 ± 11
Yakut	1.13	4.52	5.52	17 ± 15
Yukagir	3.01	2.66	4.44	9 ± 3

Table S1.4. ALDER test for admixture LD in Central Asians / Siberians. Mixe and Han Chinese are used as source populations.

2. Test of the number of Native American founding populations

To investigate whether all Native American groups from Central and Southern America^{1,4} are consistent with being derived from a single stream of ancestry, we applied $qpWave^2$ to ask the question whether the set of f_4 -statistics of the form f_4 (American₁, American₂; Outgroup₁, Outgroup₂) forms a matrix that is consistent with being of rank 0. Intuitively, if all these Native American populations descend from the same stream of migration into the Americas, then all these statistics should be consistent with 0. We test for deviations from this null hypothesis on all the f_4 -statistics jointly. We compute a single *P*-value that appropriately corrects for the correlation structure of the statistics using a Hotelling *t*-test.

The Outgroups in our analysis were 4 populations from each of 6 worldwide regions.

Africa: Yoruba, Ju_hoan_North, Dinka, Mbuti Siberia/Central Asia: Tuvinian, Mongola, Yakut, Kyrgyz East Asia: Han, Uygur, Japanese, Ami Oceania: New_Guinea, Papuan, Australian_WGA, Mamanwa South Asia: Kusunda, Onge, Kharia, Sindhi

West Eurasia: Orcadian, Spanish, Sardinian, Chechen

For Native American, we restrict to 7 groups with at least 3 unadmixed individuals.

Native Americans: Cabecar, Guarani, Karitiana, Mixe, Piapoco, Pima, Surui

We used *qpWave* to perform likelihood ratio tests for whether the matrix of statistics f_4 (American₁, American₂; Outgroup₁, Outgroup₂) is consistent with rank 0, 1, 2, or 3, which corresponds to 1, 2, 3 or 4 ancestral populations being required to explain the data. We find that 1 ancestral population was rejected ($P = 2 \times 10^{-7}$), but 2 or more were consistent with the data ($P \ge 0.09$) (Extended Data Table 1). The rejection of a single ancestral population is robust to dropping each of the 6 large geographic regions from which the Outgroups were derived ($P < 10^{-3}$), as well as dropping each of the 7 Native American populations (P < 0.01) (Extended Data Table 1).

The evidence for two ancestral populations is driven by Amazonian and Australasian populations

To determine which outgroup and Native American populations contributed the most to the rejection of rank 0, we examined the weight coefficients assigned to each population by *qpWave*. We find that the greatest coefficient among the outgroups is assigned to the Onge from the Andaman Islands. The next strongest coefficients are assigned to South Asians and Oceanians. The greatest coefficients among Native Americans are assigned to the Amazonian Suruí and Karitiana, whereas the lowest are assigned to the Central American Mixe and Pima (Extended Data Figure 2).

An alternative approach to assessing which outgroup populations contribute the most to the rejection of rank 0 is to test different subsets of the 24 outgroup populations separately. We tested different sets of 8 populations where we either paired the 4 African populations or 4 Central Asian/Siberian populations with 4 populations from another region. We find that the lowest *P*-values are found for pairings with South Asia and Oceania (Extended Data Table 1). This is contrary to what would be expected if the detected signal is due to recent gene flow between the Americas and Siberia, in which case we would expect to see the lowest *P*-values for Siberian populations. It is also contrary to what would be expected if the signal is due to recent gene flow between the signal is due to cryptic post-Columbian European or African ancestry, in which case we might expect to find the lowest *P*-values for those populations.

3. Allele frequency symmetry tests

In SI 2 we found that genome-wide data from Central and Southern Native American populations were inconsistent with a single ancestral population, and that the major inconsistencies were driven by asymmetrical affinities to South Asians and Oceanians. Within the Americas, our analyses suggested that major differences could be found between the Amazonian populations Suruí and Karitiana on the one hand, and Central American populations such as Pima and Mixe on the other.

To investigate whether there is an asymmetrical relationship to Non-American populations between Central Americans and Amazonians, we computed f_4 -statistics of the form f_4 (Chimpanzee, Non-American; Central Americans, Amazonians) where Central Americans consisted of Pima, Mixe and Kaqchikel (see SI 1) and Amazonians consisted of the Karitiana and Suruí. The f_4 -statistic⁹ is computed as the product

$$f_4(A, B; X, Y) = (p_A - p_B)(p_X - p_Y)$$

averaged over all SNPs, where p_A , p_B , p_X , and p_Y are the frequencies of an arbitrarily chosen allele in populations A, B, X and Y at each locus. Another way to think of the f_4 -statistic is as the numerator of the D-statistic¹⁰. In practice, f_4 -statistics and Dstatistics give qualitatively indistinguishable results for tests for consistency with zero¹⁰. Intuitively, the f_4 -statistic can be thought of as testing whether allele frequency differences between A and B are correlated with those between X and Y, an observation that is not consistent with a tree-like model where X and Y are from a single ancestral population. In some instances we also compute the D-statistic

$$D(A, B; X, Y) = \frac{(p_A - p_B)(p_X - p_Y)}{(p_A + p_B - 2p_A p_B)(p_X + p_Y - 2p_X p_Y)}$$

with the corresponding notation. We compute standard errors (SEs) for all statistics using a Block Jackknife weighted by the number of SNPs in each 5 cM block in the genome^{9,11}. We report *Z*-scores based on the ratio of *D*/SE. We interpret statistics |Z| > 3 as being significantly different from 0.

A genetic affinity between native Amazonians and native Oceanians

We find evidence for a significant excess in shared derived allele frequencies between Amazonians and five populations: Onge, Papuans, New Guinean highlanders, Australians from Arnhem Land, and Mamanwa Negritos from the Philippines (Z > 3). In addition, we find positive statistics for other Oceanian populations such as Bougainville Papuans, Tongans, and Ami from Taiwan, and some Indian populations such as Kharia and Bengali (Extended Data Table 2).

Robustness to different Non-American outgroups

We tested the consistency of this signal for other outgroups than Chimpanzee, replacing *A* in the test $f_4(A, B)$; Mixe, Surui) with one of Chimpanzee, Mbuti and Biaka pygmies from Central Africa, Yoruba from West Africa, Dinka from East Africa, Ju_hoan_North (San) from southern Africa, Yakut and Yukagir from Siberia, and Han Chinese. We also varied *B* among Onge, Papuans, New Guineans, and Australians. We find that the signal is highly consistent for these different combinations. In many cases it is even stronger for the non-Chimpanzee outgroups (Table S3.2).

A	В	$f_4(A, B; Mixe, Surui)$	Ζ
Chimpanzee	Onge	0.00100490	4.01
Chimpanzee	Papuan	0.00083341	3.29
Chimpanzee	New_Guinea	0.00083884	3.17
Chimpanzee	Australian_WGA	0.00085497	3.23
Mbuti	Onge	0.00088618	4.06
Mbuti	Papuan	0.00071469	3.30
Mbuti	New_Guinea	0.00072013	3.11
Mbuti	Australian_WGA	0.00073710	3.19
Biaka	Onge	0.00091768	4.36
Biaka	Papuan	0.00074620	3.56
Biaka	New_Guinea	0.00075163	3.33
Biaka	Australian_WGA	0.00076856	3.41
Yoruba	Onge	0.00085923	4.21
Yoruba	Papuan	0.00068774	3.37
Yoruba	New_Guinea	0.00069317	3.13
Yoruba	Australian_WGA	0.00071020	3.28
Dinka	Onge	0.00104014	4.90
Dinka	Papuan	0.00086865	4.14
Dinka	New_Guinea	0.00087409	3.88
Dinka	Australian_WGA	0.00089104	4.01
Ju_hoan_North	Onge	0.00102585	4.79
Ju_hoan_North	Papuan	0.00085436	3.94
Ju_hoan_North	New_Guinea	0.00085979	3.69
Ju_hoan_North	Australian_WGA	0.00087661	3.81
Yakut	Onge	0.00078838	3.95
Yakut	Papuan	0.00061689	3.21
Yakut	New_Guinea	0.00062233	2.92
Yakut	Australian_WGA	0.00063896	2.91
Han	Onge	0.00079784	4.10
Han	Papuan	0.00062635	3.34
Han	New_Guinea	0.00063179	2.95
Han	Australian_WGA	0.00064859	2.99
Yukagir	Onge	0.00090198	4.64
Yukagir	Papuan	0.00073050	3.87
Yukagir	New_Guinea	0.00073593	3.50
Yukagir	Australian_WGA	0.00075238	3.51

Table S3.2. Significant statistics of the form $f_4(A, B;$ Mixe, Surui).

Robustness to different Native American contrasts

We tested different combinations of Central American and Amazonian populations, and found consistent results for all Central Americans (Kaqchikel, Mixe and Pima), as well as for the Amazonian Karitiana and Suruí. Karitiana shows a non-significantly attenuated signal compared to the Suruí (approximately 1 SE difference) (Table S3.3).

We tested each individual of the 8 Suruí and 10 Mixe against each other in the test D(Chimpanzee, Onge; individual 1, individual 2). We find that comparisons between

two Suruí individuals are generally consistent with 0, whereas all $8 \times 2=16$ comparisons between Mixe and Suruí individuals show a positive skew. Comparisons between Mixe individuals are more dispersed, but do not show a systematic pattern. This suggests that the affinity to Onge in the Suruí is not due to sequence errors or unusual ancestry in some individuals (Extended Data Figure 3).

Α	В	Χ	Y	$f_4(\mathbf{A},\mathbf{B};\mathbf{X},\mathbf{Y})$	Z
Chimpanzee	Onge	Mixe	Surui	0.001005	4.01
Chimpanzee	Onge	Kaqchikel	Surui	0.001156	3.90
Chimpanzee	Onge	Pima	Surui	0.001089	4.01
Chimpanzee	Onge	Mixe	Karitiana	0.000631	2.79
Chimpanzee	Onge	Kaqchikel	Karitiana	0.000784	2.76
Chimpanzee	Onge	Pima	Karitiana	0.000716	2.78

Table S3.3. Different contrasts between Central Americans and Amazonians

We analyzed Native American individuals from other regions that have been genotyped on the Human Origins array, as well as individuals genotyped on Illumina arrays². The study that reported the Illumina array data masked genomic segments of post-Columbian European or African ancestry, allowing us to carry out analyses including individuals with post-Colombian mixture (and analyses restricting to individuals without any evidence of post-Columbian mixture in their genome).

We computed f_4 (Yoruba, Papuan; Mixe, Surui) in these 3 data sets, choosing Papuans as the reference Australasian population since Illumina data for the Onge have not previously been published. We find no clear evidence (|Z|>3) for any population beyond the Karitiana and Suruí as having excess affinity to Papuans in the unadmixed individuals on the Illumina array. In the ancestry masked version of the Illumina data, the Maya2 show a significant signal. While these results are intriguing, we choose not to draw strong conclusions based on the masked data, since it is possible that the local ancestry masking could affect a fine-scale signature such as the affinity between some Native Americans and Australasians that we are investigating.

Table S3.4. Generalization of findings to more populations and different datasets Affinity of Papuans to Native Americans identified as entirely of First American ancestry by Reich et al. (2012) and genotyped on Illumina and Affymetrix arrays. We present results for the statistic f_4 (Yoruba, Papuan, Mixe, Test).

	Illumina masked		Illumina unadmixed		Human Origins unadmixed	
	f_4	Ζ	f_4	Ζ	f_4	Ζ
Cree	-0.0032	-1.07				
Algonquin	-0.0050	-1.72				
Ojibwa	-0.0050	-1.89				
Pima	-0.0007	-0.41	-0.0008	-0.38	0.0001	0.31
Yaqui	0.0111	2.54				
Tepehuano	0.0011	0.75	0.0000	-0.02		
Maya1	0.0035	2.61	0.0051	1.62		
Maya2	0.0066	4.06				

Purepecha	-0.0070	-1.84				
Zapotec2	0.0020	1.38	0.0000	 0.01		••
Mixtec	0.0020	1.58	0.0000	0.01		
Zapotec1	0.0039	2.91	0.0059	 2.3		
Kaqchikel	0.0027	1.72	0.0061	1.91		
Wayuu	0.0027	1.72	0.0001	2.06	0.0031	 0.92
Kogi	0.0028	0.37	0.0049	0.53		
Arhuaco	-0.0043	-1.53	0.0014	0.55		
Maleku	0.0008	0.27	0.0011	 0.35		
Chorotega	-0.0011	-0.24		0.55		
Huetar	0.0024	0.53				••
Cabecar	0.0024	2.22	 0.0046	 2.13		 1.6
Bribri	0.0044	2.22	0.0040	2.13		
Teribe	0.0063	2.13	0.0004	2.29		
Guaymi	0.0003	2.39 1.94	0.0072	1.69		
Embera	-0.0013	-0.59	-0.0009	-0.4		
Guahibo	-0.0013	-0.39	-0.0009	-0.4		
Waunana	0.0004	-0.2 0.63	0.0001	-0.03 0.79		
Palikur	0.0013	0.83		0.79		
	-0.0019	-0.56	0.0023	-0.26	0.0002	 0.1
Piapoco Inga	0.0023	-0.36	-0.0003		0.0002	
Ticuna	0.0023	0.99	0.0026	 1.07	0.0076	 2.14
Arara	0.0023	0.99	0.0028	0.72		
Arara Parakana						
Jamamadi	0.0055 0.0005	1.67	0.0055 0.0008	1.67		
Karitiana	0.0005	0.13 1.63	0.0008	0.23 1.77		
					0.0054	
Surui	0.0075	3.15	0.0080	3.19	0.0087	3.36
Quechua	0.0019	1.29	0.0004	0.12	-0.0060	-1.83
Aymara	0.0018	1.18	0.0021	0.98	-0.0005	-0.15
Chane	0.0028	1.11	0.0018	0.68	0.0046	1.46
Wichi	0.0002	0.07	0.0018	0.74		
Guarani	-0.0002	-0.12	-0.0021	-0.93	0.0001	0.03
Kaingang	0.0008	0.27				
Toba	0.0028	1.41	0.0021	0.87		
Diaguita	0.0004	0.16				
Hulliche	0.0030	1.33				
Chilote	0.0025	1.01				
Chono	-0.0022	-0.74				
Yaghan	0.0010	0.39	0.0013	0.37		

Comparison with the Athabascan Dakelh

The three northern North American groups in the ancestry masked data showed somewhat negative statistics in Table S3.4, which could be consistent with an excess of Papuan affinity not only in South Americans but also in Central Americans, and the possibility that northern North Americans might form a better baseline for detecting excess affinities to Oceanians. However, no Northern Amerind individuals in this data set were identified as unadmixed. We therefore analyzed data from Athabascanspeaking Dakelh from Raghavan et al⁶, and used the unadmixed individuals identified in SI 1. We find no significant evidence of greater affinity to Papuans in the Pima compared to the Dakelh (Table S3.5). However we do observe Z ~2.1 for an affinity between the Dakelh and Yakut, which hints at an affinity to Siberians such as the Yakut in the Dakelh, who like the Chipewyan are also Athabascan-speakers. An affinity to Siberians would also cause an attraction between the Dakelh and Papuans, since Papuans share more genetic drift with Siberians than with Yoruba. This would

tend to diminish any signals of difference in Papuan-relatedness between the Pima and Dakelh. In summary, while we do not find any clear evidence for a difference in First American ancestry proportions in the Dakelh compared with more southern Native Americans, we also cannot exclude the possibility that northern North Americans (not Dakelh) have a lower baseline affinity to Australasians than do Central Americans.

Table S3.5. No evidence for differences in Papuan-related affinity between
unadmixed British Columbian Dakelh and Pima from Central America.

A	В	X	Y	D	Ζ
Yoruba	Papuans	Dakelh	Surui	0.0101	3.82
Yoruba	Papuans	Dakelh	Pima	0.0034	1.50
Yoruba	Papuans	Pima	Surui	0.0071	2.35
Yoruba	Yakut	Dakelh	Surui	-0.0018	-0.88
Yoruba	Yakut	Dakelh	Pima	-0.0040	-2.06
Yoruba	Yakut	Pima	Surui	0.0023	1.02

Comparison with the ancient Clovis-associated Anzick individual

We investigated the evidence for differential relatedness to Australasians using ancient Native American samples overlapped with San and Yoruba ascertained SNPs on the Human Origins array. We use Dinka as outgroup, as they are less differentiated from non-Africans than Yoruba are. This minimizes the effects of errors in the ancient attraction to the outgroup. DNA causing an We computed f_4 (Dinka, Onge/New Guinea/Papuan; Ancient, Mixe/Surui). We find no evidence for a greater affinity to Onge/New Guinea/Papuan in any ancient sample than is found in the Mixe. In contrast. we find that the Suruí show an affinity to Onge/New Guinea/Papuan compared to the ancient Anzick individual (Table S3.6).

Table S3.6.	Comparison	with the	Anzick	Clovis	sample.
-------------	------------	----------	--------	--------	---------

Z-scores greater than 2 are highlighted and only observed for comparisons with Suruí.

A	В	X	Y	$f_4(\mathbf{A},\mathbf{B};\mathbf{X},\mathbf{Y})$	Z	SNPs
Dinka	Onge	Clovis	Surui	0.000933	2.64	349,435
Dinka	New Guinea	Clovis	Surui	0.000787	2.22	349,435
Dinka	Papuan	Clovis	Surui	0.000826	2.51	349,435
Dinka	Onge	Clovis	Mixe	-0.000032	-0.10	349,435
Dinka	New Guinea	Clovis	Mixe	-0.000108	-0.33	349,435
Dinka	Papuan	Clovis	Mixe	-0.000036	-0.12	349,435

Comparisons between different Australasian groups

We tested whether there is evidence for either Oceanians or Andamanese being significantly more strongly related to Amazonians using direct contrasts of the form D(A, B; Mixe, Surui). We found that the Philippine Mamanwa show less affinity than the other populations (many |Z| > 2), consistent with their known Austronesian admixture¹². There are no consistent patterns for the other populations (Table S3.7).

				D(A, B;	
A	В	X	Y	X, Y)	Ζ
Onge	Papuan	Mixe	Surui	-0.0023	-0.85
Australian	Onge	Mixe	Surui	0.0069	2.20
Mamanwa	Onge	Mixe	Surui	0.0069	2.76
Australian_WGA	Onge	Mixe	Surui	0.0020	0.67
New_Guinea	Onge	Mixe	Surui	0.0022	0.77
Australian	Papuan	Mixe	Surui	0.0052	2.06
Mamanwa	Papuan	Mixe	Surui	0.0048	2.00
Australian_WGA	Papuan	Mixe	Surui	-0.0003	-0.15
New_Guinea	Papuan	Mixe	Surui	-0.0001	-0.08
Australian	Mamanwa	Mixe	Surui	-0.0002	-0.07
Australian	Australian_WGA	Mixe	Surui	0.0059	2.33
Australian	New_Guinea	Mixe	Surui	0.0055	1.95
Australian_WGA	Mamanwa	Mixe	Surui	-0.0050	-1.83
Mamanwa	New_Guinea	Mixe	Surui	0.0048	1.78
Australian_WGA	New_Guinea	Mixe	Surui	-0.0003	-0.11

Table S3.7. Direct comparisons between Andamanese and other Oceanians.

Consistency of the link between Australasians and Amazonians in different data We assessed whether the affinity of Amazonians to Australasians was robust in Human Origins data as well entirely genetic data sets generated using different technological platforms. We use f_4 (Yoruba, Papuan; Mixe, Surui) throughout since these four populations are present in each of the datasets . In Table S3.8 we list this statistic for Illumina SNP arrays, Affymetrix Human Origins (HO) SNP arrays, and Illumina high-coverage sequencing. For all these data we find evidence of significant (Z>3) affinity between the Suruí and Papuans.

	f_4	Ζ	SNPs	n _Y	n _P	$\mathbf{n}_{\mathbf{M}}$	$\mathbf{n}_{\mathbf{S}}$
Illumina masked	0.000776	3.15	364,428	21	16	17	24
Illumina unadmixed	0.000823	3.21	364,470	21	16	9	24
SGDP genomes	0.001265	4.00	9,873,045	3	16	3	2
Affymetrix HO	0.000688	3.37	593,142	70	26	10	8

Table S3.8. f_4 (Yoruba, Papuan; Mixe, Surui) in different data sets. n_Y , n_P , n_M and n_S refer to the sample size of Yoruba, Papuans, Mixe, and Suruí in each data set.

Consistency of the signal for different mutation classes

We stratified the *D*-statistics for the complete genome data into each separate nucleotide substitution class, and found highly consistent results for all 6 classes (within 2 SEs) (Table S3.9). This suggests that the results are not due to convergent evolutionary processes, for example, correlations in the effectiveness of gene conversion on the lineages leading to Suruí and Papuans.

Mutation class	1		Informative SNPs
A / T	0.0169	2.63	60,538
A / G	0.0191	3.64	268,962
A / C	0.0208	3.49	67,210
G / T	0.0248	4.27	67,623
C / T	0.0220	4.24	270,133
C / G	0.0248	4.26	64,951

Table S3.9. Stratification of D(Yoruba, Papuan; Mixe, Surui) by mutation class.

These analyses are performed on the full genome sequencing data.

Consistency of the signal of relatedness between Australasians and Amazonians for different ascertainment schemes

We studied all 13 different ascertainment panels that underlie the full Human Origins array SNP set. Some of these panels contain relatively few SNPs. However, we find strong statistics for ascertainments as different as Han Chinese and Yoruba, and no evidence for any one panel contributing disproportionally to the signal (Table S3.10).

Table S3.10. An affinity between Suruí and Papuans is seen across multiple ascertainment schemes of the Human Origins array. Rows are ordered by the number of SNPs analyzed (more SNPs gives more precision).

Ascertainment individual	<i>f</i> ₄ (Yoruba, Papuan; Mixe, Surui)	Z	SNPs (autosomal)
Union of 13 panels	0.000688	3.37	593,142
San	0.000525	2.24	156,365
Denisova-San differences	0.000611	2.81	140,044
French	0.000478	1.13	108,311
Yoruba	0.000894	3.16	119,765
Han	0.001587	3.27	75,390
Papuan1	0.001009	2.14	46,676
Cambodian	0.00026	0.40	16,442
Melanesian	0.001247	1.82	14,449
Sardinian	0.000843	1.16	12,470
Mbuti	0.001186	2.34	11,745
Papuan2	0.001753	2.30	11,720
Mongolian	-0.000546	-0.65	10,389
Karitiana	0.000003	0.00	2,555

Admixture signals as a function of proximity to functionally important regions

To further characterize the excess genomic affinity between Amazonians and southeast Asian populations, we divided the genome into 10 deciles of the '*B*-value' proposed by McVicker et al.¹³ which integrates multiple genomic annotations into a single score for functional importance for each base pair in the genome. We computed D(Yoruba, Papuan; Mixe, Surui) separately for each bin. We observe that the bin that is closest to functionally important elements shows the most asymmetry ($D = 0.0397 \pm 0.0197$) and than the bin that is most distant from functional elements shows the least ($D = 0.00837 \pm 0.0106$) (Extended Data Figure 4A). We fit a linear regression to D as a function of B, and estimate the slope to be -0.0004.

To compute standard errors, we used a weighted Block Jackknife procedure where one 5 Mb block of the genome is dropped in turn and the linear regression is recomputed. The variability of this statistic can be used to obtain an estimate of the standard error^{9,11}, which we weighted using the number of informative loci in each block. We estimate the *Z*-score for the linear coefficient being different from zero to be -1.95 (one-tailed *P*-value = 0.026). To test if this observation is independent from the observation of a genome-wide significant *D*-statistic, we implemented a twodimensional jackknife for both the linear coefficient and the *D*-statistic.

To understand the expected behavior of the correlation between *B* and *D* for known signals of asymmetry, we examined the relationship between *B* and *D* for a larger number of population comparisons. For this analysis, we used previously published genomes from Yoruba, San, Mbuti, Han, Dai, Sardinians, French, Australians, Papuans, Karitiana and Mixe^{14,15}, together with the 18 newly reported genomes from Yoruba, Papuans, Mixe and Suruí reported in this study. We computed all possible linear coefficients for *D* as a function of *B* for all 495 quartets (Yoruba, X; Y, Z). We found that all quartets of populations for which the *Z*-score for the slope was significant (|Z| > 3) also showed a significant genome-wide *D*-statistic with the opposite sign (Extended Data Figure 4B).

We hypothesize that the explanation for this phenomenon is that allele frequencies in isolated populations become more differentiated in the vicinity of functionally important regions due to linked selection imposing increased evolutionary stochasticity. This emulates genetic drift (or a low effective population size), which makes admixture between more differentiated populations easier to detect in the sense that the magnitude of allele frequency skews that are used by statistics such as D is increased. This suggests that genome-wide signals of admixture in modern humans are systematically stronger near functional regions, but does not imply that ancestry itself is systematically depleted in these regions.

Having validated this test, we applied it to our signal of interest. This yielded a P-value of 0.00014, which is in fact less significant (larger P-value) than the genomewide D-statistic alone. This suggests that the monotonic relationship between D and Bdoes not provide any statistical evidence for admixture above and beyond that of the genome-wide D-statistic alone. Thus, the value of these results is not to increase the strength of the signal, but rather to show that the direction of the signal is what is expected for a real biological effect.

Analysis of 9 newly genotyped populations from Brazil

We also performed allele frequency based symmetry specifically using the 9 newly genotyped Brazilian populations. We find that the new Suruí and Karitiana data both show strong signals with African Yoruba as outgroup. When we use Han Chinese as outgroup, the Xavante also show a strong signal (Z = 3.25) and all Amazonians except the Urubu_Kaapor show moderate signals (Z > 2) (Table S3.11).

Table 55.11. Animity to Onge in 9 newly genotyped brazinan populations							
	D(Yoruba, On	ge; Mixe, X)	D(Han, Ong	ge; Mixe, X)			
X	D	Z	D	Ζ			
Apalai	0.0037	1.55	0.0066	2.92			
Arara	0.0043	1.49	0.0072	2.64			
Guarani_GN	0.0021	0.74	0.0078	2.96			
Guarani_KW	0.0032	1.35	0.0060	2.66			
Karitiana	0.0082	3.10	0.0107	4.14			
Suruí	0.0092	3.27	0.0084	3.01			
Urubu_Kaapor	0.0042	1.49	0.0037	1.34			
Xavante	0.0044	1.98	0.0071	3.25			
Zoro	0.0052	1.37	0.0099	3.24			

Table S3.11.	Affinity to O	nge in 9 new	ly genotyped	Brazilian n	opulations
	i i i i i i i i i i i i i i i i i i i	nge m / new	i Senocypea	Di azinan p	opulations

We find no significant difference in affinity to the Onge comparing the new Karitiana and Suruí genotypes obtained from blood samples to those obtained from Human Genome Diversity Project (HGDP) cell lines. This is an important observation, as it shows that our signal cannot be a cell line artifact (Table S3.12).

Table S3.12. The signal is not a cell line artifact

A	В	X	Y	D(A, B; X, Y)	Ζ
Yoruba	Onge	Surui _{blood}	Surui _{cell_line}	-0.00209	-0.92
Yoruba	Onge	Karitiana _{blood}	Karitiana _{cell line}	0.00039	0.21
Han	Onge	Surui _{blood}	Surui _{cell_line}	-0.00242	-1.11
Han	Onge	Karitiana _{blood}	Karitiana _{cell line}	0.00293	1.66

4. Linkage disequilibrium symmetry tests

We devised a novel linkage disequilibrium statistic that measures symmetry in linkage disequilibrium between two proposed clades with a pair of populations in each. The statistic, which we refer to as h_4 , is:

$$h_4 = \left((p_{12}^A - p_1^A p_2^A) - (p_{12}^B - p_1^B p_2^B) \right) \times \left((p_{12}^C - p_1^C p_2^C) - (p_{12}^D - p_1^D p_2^D) \right)$$

where 1 and 2 are arbitrarily chosen references alleles at two different loci, respectively, and A, B, C, and D denote four different populations. Thus, p_{12}^A is the frequency of the 12 haplotype in population A, and p_1^A is the frequency of the 1 allele in population A. The quantity $H_{12}^A = p_{12}^A - p_1^A p_2^A$ measures the difference between the observed haplotype frequency and the expected haplotype frequency given the allele frequencies¹⁶, and corresponds to the classical population genetic quantity "D" (this should not be confused with the D-statistic used to test for consistency with a tree elsewhere in this study).

The motivation for this statistic being informative about population history is that under a tree-like model ((A, B), (C, D)), differences in linkage disequilibrium between populations *C* and *D* are not expected to be correlated to differences in LD between populations *A* and *B*. If there has been gene flow between the two clades however, the statistic may be significantly positive or negative, much like f_4 and *D*-statistics⁴.

In practice, we compute this statistic for each polymorphic locus ('target locus') by identifying all other polymorphic loci 5' of the target locus in a window extending w cM from a focal point at distance interval d. We average the statistic over all valid pairs of loci in the genome identified in this way. We compute standard errors by a Block Jackknife over contiguous 0.5 cM blocks, where SNP pairs that bridge the boundary of two blocks are assigned to the block in which the target locus is found.

Test simulation

To test the h_4 statistic, we simulated a history modified from Lipson et al¹⁷. We simulated 500 chromosomes of 50 kb each. At $0.02 \times 4N_e$ generations ago, 60% of the ancestry of population 6 was contributed by population 5 and the remainder by population 3. We ascertained polymorphisms in 'pop1', made up of 2 chromosomes from pop2 that were not used in h_4 computations. The *ms* command line was

'Base model':

ms 254 500 -t 500 -r 500 50000 -I 7 2 50 50 50 50 50 2 -ej 0.0 2 1 -es 0.02 6 0.4 -ej 0.06 6 3 -ej 0.04 8 5 -ej 0.08 5 4 -ej 0.12 4 3 -ej 0.2 3 1 -ej 0.3 1 7 -en 0.3 7 1

We also simulated a bottleneck scenario where the effective population size of population 5 was reduced by a quarter. The following command was added to the above command line: '-en $0.05 \ 0.25$ -en $0.04 \ 5 \ 1$ '

Model	NO admixture		Admixture	
	phased	unphased	phased	unphased
Basic model	0.185	0.354	9.475	5.689
Bottleneck in pop5	-1.459	-1.739	7.124	3.368

Table S4.1. Simulation results from applying the h_4 (pop2, pop3; pop4, pop5) statistic for SNP pairs within a distance of 10,000 bp. The numbers are Z-scores.

We computed the h_4 -statistic for all SNP pairs within 10,000 base pairs of each other both for the perfectly phased simulated data as well as for versions where the phase was randomized in pairs of 2 chromosomes (to mimic unphased human SNP data). The h_4 -statistic finds significant evidence of gene flow when testing populations that were involved in the admixture event, with the signal still present (albeit weakened) when phase information is scrambled (Table S4.1). As we would hope for a useful test, sets of populations that are related according to a simple tree give non-significant statistics.

Application to data from the Human Origins array

We phased individuals typed on the Human Origins array panel 5 (119,765 autosomal SNPs ascertained in a Yoruba individual) using SHAPEIT with default parameters (SI 1). We then computed h_4 (Yoruba, B; X, Y) with B, X and Y drawn from the populations: Australian, Dai, French, Han, Ju_hoan_North, Karitiana, Mbuti, Mixe, Onge, Papuan, Pima, Sardinian, Suruí, and Yakut. We find that D and h_4 are largely highly correlated, but that there are also a few cases where they give conflicting significant results (Extended Data Figure 5C). When all Africans and Oceanians (Mbuti, San, Papuan, Australian) are removed, there are no such conflicts (Extended Data Figure 5D). One possible explanation is that these 4 populations are the only ones that harbor ancestry that is basal to Yoruba (which we have used to ascertain SNPs). The h_4 -test thus does not seem be appropriate when populations used in the test branched off prior to the population(s) used in SNP ascertainment. However the significant h_4 -statistic we observe in the Onge as described below cannot be explained by this bias.

We computed h_4 -statistics of the form h_4 (Yoruba, X; Mixe, Surui) for all populations X in the Human Origins array, and all pairs of SNPs within 0.01 cM of each other. We restrict the analysis to populations with at least 6 individuals. We find support for an affinity between Oceanians and Onge to Suruí, with larger Z-scores than is the case for the standard *D*-statistic based on allele frequencies. Four non-African populations, including the Onge and native New Guineans and Papuans, show Z > 3 (Table S4.2). We also computed the h_4 -statistic for windows of 0.001 cM centered around different genetic distances for selected populations (Extended Data Figure 5E). We find that the signal dissipates at approximately 0.02 cM.

	Population	h_4	SE	Z	Loci	N	Region
1	New_Guinea	0.0004195	7.3E-05	5.71	14938	38	Oceania
2	Onge	0.0003474	7.8E-05	4.43	14938	22	S. Asia
3	Papuan	0.0003193	6.9E-05	4.60	14938	52	Oceania
4	Bougainville	0.0002452	7.2E-05	3.43	14938	20	Oceania

Table S4.2. Significant statistics for h_4 (Yoruba, non-African; Mixe, Surui).

The most negative statistics in non-Americans are found for Northeast Asians and Siberians (*e.g.* Z = -2.6 for Evens), which would be expected if the other founding population of the Americas (the population without the strong affinity to Austrasians) was related to present-day Siberians (Figure 1B). This is as expected for conventional models for the ancestry of First Americans.

Caveats

One qualitative feature that differs between f_4 -statistics and h_4 -statistics is that h_4 statistics do not have the Martingale property in relation to genetic drift. Specifically, the h_4 -statistic can be biased by different degrees of genetic drift since divergence.

For example, for $h_4(A, B; X, Y)$, if Y has more SNP pairs without polymorphism than X, this will in turn result in more SNP pairs with $H_{12}^Y = 0$ than $H_{12}^X = 0$. Thus we might expect that if B also has many SNP pairs with $H_{12}^B = 0$, we could see falsepositive genome-wide h_4 -statistics indicating affinity between B and Y. To assess the impact of this in our empirical data, we estimated the fraction of SNP pairs with H_{12} = 0 in all populations. Extended Data Figure 5A shows the fraction of SNP pairs with $H_{12} = 0$ on the x-axis and h_4 (Yoruba, Test; Mixe, Surui) on the y-axis. Papuans, New Guineans and Onge are clearly outliers in that they show a strong affinity to Suruí, like the Amazonian Karitiana. We also see significant statistics for Africans, but we note that we have ascertained in Yoruba for this analysis, and so it is not valid to use populations that may have ancestry basal to Yoruba for the purpose of this test since it breaks the assumption of polymorphism in the ancestral population. We see that there are several East Asian and Siberian populations that have similar fractions of SNP pairs with $H_{12} = 0$ as the populations with significant h_4 -statistics. However, these East Asians and Siberians do not show significant evidence of attraction to the Suruí. This suggests that the fraction of SNP pairs with D = 0 is not severely impacting the significant h_4 -statistics that we are detecting.

5. Chromosome painting symmetry tests

We used SHAPEIT to phase 593,142 SNPs with the same set of individuals as described above along with the 48 newly genotyped Brazilian individuals, using all panels of the Human Origins array. We 'painted' the chromosomes of unadmixed Native American individuals using non-American populations as donors, but excluded the Yukagir and the Chukchi since they have evidence of back-migration from the Americas. We ran CHROMOPAINTER v2 using default parameters, painting each recipient individual separately, but using all donor populations as candidates to paint each recipient haplotype. To assess statistical uncertainty, we repeated this procedure for each recipient individual using 22 subsets of the data where for each separate subset a different chromosome had been dropped. We then used the results of these 22 block jackknife pseudoreplicates to obtain a weighted Block Jackknife estimate of the standard error for our test statistic (see below).

To test if the recipient populations copied equally from the donor populations, we computed the average chunk count $C_{R:D}$ copied from a given donor population D in each recipient population R (averaged over individuals). We then computed a $S(R_1, R_2; D)$ statistic that quantifies the symmetry between two Native American populations in their copying from each donor population

$$S(D; R_2, R_1) = \frac{C_{R_1:D} - C_{R_2:D}}{C_{R_1:D} + C_{R_2:D}}$$

If two Native American populations, such as the Suruí and the Mixe, derive all of their ancestry from a single ancestral population, we expect that they would copy from the donor populations at an equal rate. We computed the standard error of this statistic using the 22 subsets of the data where each autosome had been dropped, weighted using the number of SNPs on each chromosome.

We fixed R_2 =Mixe and computed all combinations of non-Americans other than Yukagir and Chukchi as *D* and Native Americans other than Mixe as R_1 . Table S5.1 lists all top *Z*-scores for the *S*-statistics obtained for each population R_1 , and we find that the great majority of Native American populations with a *Z*-score of 3 or greater show an Australasian population as maximizing this symmetry statistic (Onge, Australian, Bougainville, New_Guinea or Tongan). The exceptions are Chane, Zoro and Pima. The Chane shows a highly significant rate of copying from Turkish_Jew and other European populations such as Norwegians, suggesting that this single Chane individual might have cryptic European ancestry. Similarly, the single Zoro individual show a top *Z*-score for African Wambo. We caution against strong interpretations of this since single individuals represent these two groups. The Pima show an affinity to African Tshwa (Z = 3.71), possibly suggesting cryptic African ancestry. In Table S5.2 we show statistics for all Native American populations when Onge is used as donor, which is the statistic underlying the heatmap in Figure 1D.

R ₁	D	$S(D;R_2,R_1)$	Z
Karitiana_UFRGS	Australian_WGA	0.0059	5.58
Ticuna	Bougainville	0.0028	3.44
Surui	Onge	0.0027	4.86
Chane	Turkish_Jew	0.0027	6.21
Karitiana	Onge	0.0027	5.04
Wayuu	New_Guinea	0.0027	3.19
Surui_UFRGS	Onge	0.0026	5.26
Zoro	Wambo	0.0023	3.73
Urubu_Kaapor	Tongan	0.0023	4.31
Xavante	Onge	0.0022	4.27
Piapoco	Onge	0.0019	3.09
Guarani_KW	New_Guinea	0.0018	2.95
Arara	Onge	0.0017	2.76
Guarani_GN	New_Guinea	0.0016	2.26
Cabecar	Onge	0.0016	3.61
Guarani	Onge	0.0016	3.19
Bolivian	Kinh	0.0016	2.76
Pima	Tshwa	0.0014	3.71
Aymara	Tshwa	0.0012	2.24
Mayan	Turkmen	0.0010	2.73
Apalai	Gui	0.0009	2.76

Table S5.1. Donor populations that maximize the excess copying in a Native American population when Mixe is used as a baseline.

Kaqchikel	Ukrainian	0.0007	2.65
Quechua	Egyptian	0.0007	2.14

Table S5.2. Symmetry statistics for haplotype copying from the Onge using Mixe
as baseline for diverse Native American populations

Χ	S(Onge; Mixe, X)	Z
Karitiana_UFRGS	0.00295	4.25
Surui	0.00274	4.86
Karitiana	0.00270	5.04
Surui_UFRGS	0.00260	5.26
Zoro	0.00232	2.06
Xavante	0.00222	4.27
Piapoco	0.00189	3.09
Chane	0.00176	2.31
Arara	0.00175	2.76
Cabecar	0.00162	3.61
Guarani	0.00162	3.19
Guarani_KW	0.00151	2.76
Urubu_Kaapor	0.00147	1.97
Apalai	0.00142	2.04
Aymara	0.00119	1.16
Guarani_GN	0.00111	1.60
Bolivian	0.00105	1.59
Ticuna	0.00098	0.85
Pima	0.00084	1.74
Quechua	0.00083	0.72
Chipewyan	0.00073	1.42
Mayan	0.00062	0.66
Wayuu	0.00010	0.11
Kaqchikel	-0.00008	-0.13

6. Models of population history

Our analyses suggest that Amazonian groups such as Karitiana and Suruí share more derived alleles with Oceanian aboriginal groups and Negritos than with other Native Americans from Central and South American. This suggests that the history of Amazonians and other Americans cannot be accurately described as a simple tree. To investigate which possible alternative models of population history could fit the data, we used an admixture graph framework to test formally different hypotheses.

We used ADMIXTUREGRAPH^{4,9} to fit suggested phylogenies with admixture events to the data, and assessed goodness-of-fit by investigating all possible f_4 -statistics predicted by the fitted model and assessing whether they differed significantly from

the empirically observed statistics. We chose as a starting point the model relating Mbuti Africans, Onge, MA1 and Karitiana found by Lazaridis et al¹ where MA1 is basal to the other non-Africans but contributed ancestry to the ancestral population of Karitiana. We added to this Han Chinese as being more closely related to the ancestral population of Native Americans than the Onge are, as well as Suruí as a sister group to Karitiana and Mixe (Extended Data Figure 6A). We find that this model is inconsistent with the data, in line with the previously reported results in SI 2 and SI 3, since it predicts that Mixe and Suruí/Karitiana are equally related to Onge, and indeed we observe several statistics for which the Z-score for the difference between the predicted and empirical statistics is |Z| > 3 (Extended Data Table 3). To account for this, we fitted a model in which the ancestors of Amazonians received admixture from a population related to the Onge (Extended Data Figure 6B), and found that this provided an excellent fit to the data, with no |Z|-scores greater than 3.

To investigate if alternative admixture models could explain the data, we tested a model in which the gene flow is instead from a population closer to Han Chinese than the Onge, into Amazonians or Central Americans (Extended Data Figure 6C). This model predicts several statistics that are inconsistent with the data (Extended Data Table 3), such as the observation that Amazonians are closer to the Onge than Han Chinese (*e.g.* f_4 (Han, Onge; Mixe, Surui) >> 0). Finally, we tested a model in which the Mixe received admixture from an East Asian source more closely related to the Han (Extended Data Figure 6D), or an ancient Siberian source most closely related to mA1 (Extended Data Figure 6E), but found that these models were also unable to reproduce the empirical evidence of the Amazonians being closer to Onge.

Plausible range of the admixture fraction related to Australasians

The topology of the admixture graph that we infer makes it impossible to use a measure such as an f_4 -ratio to infer the proportion of ancestry related to Australasians and obtain confidence intervals. Instead, we tested different proportions of this ancestry in the Suruí, assuming that the proportion in Mixe is 0, and determined the proportions for which the maximum Z-score between predicted and observed f_4 -statistics is less than 3. We find that the plausible range of Australasian-related ancestry in the Suruí is 0.7-9.8% and that the lowest maximum Z-score is obtained for an admixture proportion of 2.3% (Extended Data Figure 7). If we instead find the proportion for which no Z-score is greater than 2, we obtain a range of 1.5-5.7%.

We also fitted models where the Mixe received Australasian-related gene flow. We fixed the proportion of this ancestry in the Mixe, while allowing ADMIXTUREGRAPH to fit a secondary and larger proportion in the Suruí. We find that a mixture proportion of 2.3% or more the Mixe results always produces |Z|-scores larger than 3, and so we can rule out proportions of Australasian-related ancestry in the Mixe larger than this. In the model where Mixe have 2.3% Australasian-related ancestry, the Suruí are fitted as having an extra 1.0% such ancestry, above and beyond

that in the Mixe (Extended Data Figure 7). Thus, even if the Mixe, too, have Australasian related ancestry, we can put a stringent upper bound on it.

Fitting models in which Population Y is itself admixed

We added the Africa Dinka as an additional outgroup to the Admixture Graph. We also added the Mesoamerican Pima, the northernmost population in our data with no evidence of being other than 100% First American in previous studies. We finally added in the Amazonian Xavante ,which were newly genotyped for this study.

We used this extended Admixture Graph model to explore scenarios in which the Australasian related ancestry in Amazonians derived from a Native American founding "Population Y" that was admixed with a First American lineage at the time that it contributed to the ancestors of Amazonians. While we have no statistical support for such a model, the motivation for exploring it is that from a human migration point of view, it seems plausible. It seems unlikely that population sharing 100% of its ancestry with the Onge reached South America without admixing with other populations who were inhabiting the same region.

To explore this family of models, we modeled Population Y as deriving a proportion α of their ancestry from the Onge-lineage and $(1-\alpha)$ from a basal First American lineage. We then allowed for the common ancestral population of the three Amazonian populations to carry a proportion γ from the Population Y lineage and a proportion $(1-\gamma)$ from a Mesoamerican population more closely related to the Mixe than the Pima. Figure 2A illustrates this model. We fitted admixture graphs for $\gamma = 0$ -100% and $\alpha = 0$ -100% (with a grid size of 1%) and used the number of f_4 -statistics predicted by the model that deviated by more than 3 sigma from the empirical statistics to evaluate model fit. We also fitted γ and α automatically and obtained point estimates of $\alpha=2\%$ and $\gamma=63\%$, but find that a much broader range of parameter combinations are consistent with the data. These results show that the proportion of Population Y ancestry in Amazonians can plausibly be quite high: indeed, as high as 63% or higher.

Importantly, while the proportion of Population Y ancestry in Amazonians is poorly determined by our analyses, the proportion of ancestry related to the Onge is tightly constrained. In our model, the proportion of Onge-related ancestry in Amazonians is the product of γ and α , which our model fitting shows to be constrained between 1% and 2% (no parameter combinations with a proportion outside this range fitted the data). This is similar to the estimates we obtained with modeling that used fewer populations and assumed Population Y to be an unadmixed sister group of the Onge.

Supplementary References

- 1 Lazaridis, I. *et al.* Ancient human genomes suggest three ancestral populations for present-day Europeans. *Nature* **513**, 409-413, doi:10.1038/nature13673 (2014).
- 2 Reich, D. *et al.* Reconstructing Native American population history. *Nature* **488**, 370 374 (2012).
- 3 Wang, S. *et al.* Genetic Variation and Population Structure in Native Americans. *PLoS Genet* **3**, e185, doi:10.1371/journal.pgen.0030185 (2007).
- 4 Patterson, N. *et al.* Ancient admixture in human history. *Genetics* **192**, 1065-1093 (2012).
- 5 Alexander, D., Novembre, J. & Lange, K. Fast model-based estimation of ancestry in unrelated individuals. *Genome Res* **19**, 1655 1664 (2009).
- 6 Raghavan, M. *et al.* The genetic prehistory of the New World Arctic. *Science* **345**, doi:10.1126/science.1255832 (2014).
- 7 Li, J. *et al.* Worldwide human relationships inferred from genome-wide patterns of variation. *Science* **319**, 1100 1104 (2008).
- 8 Loh, P.-R. *et al.* Inference of admixture parameters in human populations using weighted linkage disequilibrium. (2012).
- 9 Reich, D., Thangaraj, K., Patterson, N., Price, A. L. & Singh, L. Reconstructing Indian population history. *Nature* **461**, 489-494 (2009).
- 10 Green, R. E. *et al.* A Draft Sequence of the Neandertal Genome. *Science* **328**, 710-722 (2010).
- 11 Busing, F. M., Meijer, E. & Van Der Leeden, R. Delete-m jackknife for unequal m. *Statistics and Computing* **9**, 3-8 (1999).
- 12 Reich, D. *et al.* Denisova Admixture and the First Modern Human Dispersals into Southeast Asia and Oceania. *The American Journal of Human Genetics* **89**, 516-528, (2011).
- 13 McVicker, G., Gordon, D., Davis, C. & Green, P. Widespread genomic signatures of natural selection in hominid evolution. *PLoS genetics* **5**, e1000471 (2009).
- 14 Meyer, M. *et al.* A High-Coverage Genome Sequence from an Archaic Denisovan Individual. *Science* **338**, 222-226, doi:10.1126/science.1224344 (2012).
- 15 Prufer, K. *et al.* The complete genome sequence of a Neanderthal from the Altai Mountains. *Nature* **505**, 43-49, doi:10.1038/nature12886 (2014).
- Robbins, R. B. Some applications of mathematics to breeding problems III. *Genetics* 3, 375 (1918).
- 17 Lipson, M. *et al.* Efficient Moment-Based Inference of Admixture Parameters and Sources of Gene Flow. *Molecular Biology and Evolution* **30**, 1788-1802, doi:10.1093/molbev/mst099 (2013).